

**STIMULATION AND SUBSTITUTION
OF BONE ALLOGRAFT AROUND NON-
CEMENTED IMPLANTS**

Ph.D. thesis

Thomas Bo Jensen

Faculty of Health Sciences

University of Aarhus

2003

Stimulation and substitution of bone allograft around non-cemented implants

Abbreviations	4
Abstract	6
Introduction	7
Background	8
Grafting in reconstructive surgery of the hip	8
Biomechanics of impacted morselized bone allograft	9
Incorporation of impacted bone allografts	10
Risks and disadvantages associated with the use of bone graft	9
Bone graft substitutes based on osteoconductivity	15
Bone growth factors and implant healing	12
Enhancement of bone grafts and osteoconductive bone substitutes	15
Materials and methodological considerations.....	17
Ethical considerations	17
Animals	17
Observation time	17
Implants.....	17
Experimental model	17
Design	18
Grafting materials	18
Mechanical evaluation	20
Histomorphometry	21
Reproducibility.....	21
Statistics	22
Results.....	22
Exclusions	22
Study II.....	23
Study III	25
Study V	26
Discussion	27

Stimulation and substitution of bone allograft around non-cemented implants

This thesis is based on the following papers and manuscripts:

I: Jensen-TB, Overgaard-S, Rahbek-O, Lind-M, Bünger-C, Søballe-K. Osteogenic Protein-1 Device® Increases Bone Graft Resorption and Bone Formation Around Cementless Implants. *Acta Scandinavica Orthopaedica*, 2002, 73(1) p31-39

II: Jensen-TB, Overgaard-S, Rahbek-O, Lind-M, Bünger-C, Søballe-K. Osteogenic Protein-1 Device® Increases Fixation of Implants Grafted with Morsellized Bone Allgraft and ProOsteon. An experimental study in dogs. Resubmitted *J. Bone Joint Surg. (Br)*, 2002

III: Jensen-TB, Overgaard-S, Rahbek-O, Lind-M, Bünger-C, Søballe-K. The influence of OP-1 d Device on Mechanical Properties of Impacted Morsellized Bone Allograft after 3 Weeks. Manuscript 2003

IV: Jensen-TB, Overgaard-S, Rahbek-O, Søballe-K, The Influence of Platelet Rich Plasma on Osteointegration of Fresh Frozen and Processed Bone Allograft. Submitted *J. Arthro.* 2002

V: Jensen-TB, Overgaard-S, Rahbek-O, Søballe-K, Does Platelet Rich Plasma Influence Fixation of Non-cemented Implants. Resubmitted *J. Orthop. Res.* 2003

Abbreviations

AGF: Autologous growth factor

BMP: Bone morphogenic protein

EGF: Epidermal growth factor

FGF: Fibroblast growth factor

HA: Hydroxyapatite

IGF: Insulinlike growth factor

OP-1: Osteogenic protein-1

PC: Platelet concentrate

PDGF: Platelet derived growth factor

PRP: Platelet rich plasma

TCP: Tricalciumphosphate

THA: Total hip arthroplasties

Ti-6Al-4V: Titanium-6 aluminium-4 vanadium

Definitions

Bioactive materials: Materials which elicit or modulate biological activity

Biocompatible: The ability of a material to perform with an appropriate host in a specific application

Cytokine: Poly peptide regulator of cell-to-cell interaction in relation to the immunological system

Graft:

-autograft: Bone tissue harvested from and implanted in the same individual

-allograft: Bone tissue harvested from one individual and implanted in another individual, same species

-Xenograft: Harvested from one species and implanted in another

Growth factors: Polypeptides which act as signalling agents for cells

Histomorphometry: Quantitative evaluation of tissue

Implant: A device made of a biomaterial that is intentionally placed in the body covered totally or partly by an epithelial surface

Implant in- or ongrowth: Bone coverage of the implant surface in percentage

Bone incorporation: Integration of a biomaterial by bone

Osteoconduction: A process where tissue involved in bone formation is lead conducted on the surface of the biomaterial

Osteogenesis: Local bone formation occurring when bone forming cells are transplanted from one site to another

Osteoinduction: Biochemical stimulation of new bone formation at an ectopical place

Resorption: Reduction of a material due to dissolution or cellular activity

Shear: Force or stress occurring under displacement of two parallel surfaces

Strain: Relative deformation of an object

Stress: The forces that develop within a material when external load is applied

Preface

This thesis is based on experimental studies performed at Orthopaedic Research Group, Department of Orthopaedics, Aarhus University Hospital during my employment as a Diploma student in 1997 (grant from Aarhus University) and during my enrolment as a research fellow in 1999-2003 at the Department of Orthopaedics, Amtssygehuset, Aarhus University Hospital (grants from Danish Rheumatism Association and Aarhus University).

All surgical procedures and animal handling was done at the facilities of the Institute of Experimental Clinical Research. Preparation and sectioning of tissue and following evaluation was done at Orthopaedic Research Lab.

My supervisors were professor Kjeld Søballe, M.D., D.M.Sc., professor Søren Overgaard, M.D., D.M.Sc. and professor Cody Bünger, M.D., D.M.Sc. who took valuable time of to solve problems and kindly commented my work. A special thank to Søren Overgaard for support during the operationspending weekends in the operation room.. I owe specialA sincere gratitude is expressed toto Kjeld Søballe for his astonishing enthusiasm and his involving me in studies and arranging my stay at theintroducing me to Orthopaedic Biomechanics Laboratory, Hennepin County Medical Center, Minneapolis, MN, US in 2001-2002. A special thank is directed to Joan Bechtold, Ph.D. for comments to the manuscripts.

Professor Fleming Melsen, M.D., D.M.Sc. is thanked for is friendly attitude and his expertise in the evaluation of the bone-implant specimen.

My co-workers Ole Rahbek, M.D., Ph.D. and Martin Lind, M.D., Ph.D., D.M.Sc. are thanked for good assistance and comments to the articles.

These studies could not have been done without the knowledge and skills of lab technicians Anette Milton and Jane Pauli.

Acknowledgements

The studies were financially supported by The Danish Rheumatism Association, the Danish Medical Research Council, Institute of Experimental Clinical Research, the Velux Foundation, Beckett foundation and the Aarhus University.

The following companies contributed with materials:

Biomet: Implants

Stryker Biotech: OP-1 device

Interpore-Redcross: ProOsteon 200

Abstract

Bone grafting is important in the revision of failed total joint replacements. However the clinical results on revisions still do not match those of the primary inserted arthroplasties. Histological examinations of impacted bone in revision surgery often show incomplete incorporation and significant subsidence of the prosthesis is often seen. One aim of the thesis was to investigate if bone growth factor osteogenic protein-1 (OP-1) or platelet rich plasma (PRP) increases fixation and bone incorporation of bone allografted implants.

Bone allograft, especially fresh frozen, is associated with a number of potential disadvantages and risks however. Risks of viral transmission, bacterial contamination and insufficient supply might limit the use of bone allograft. Different processing techniques almost eliminate the risk of viral transmission or bacterial contamination. Substitutes such as granular ceramics might replace or expand the volume of bone allograft in the future.

The thesis is based on five experimental *in vivo* studies in canines. In all studies, we used non-loaded, stable, hydroxyapatite (HA) coated implants surrounded by a gap. Observation time was three weeks. Evaluation was based on histomorphometry and mechanical push-out tests.

In study I and II we examined the effect of recombinant human OP-1 in combination with bone allograft, HA granules or a composite of those. We showed, that OP-1 not only stimulates bone formation but also accelerated bone graft resorption dramatically. As a consequence, OP-1 did not increase fixation of bone allografted implants. Since ProOsteon is very slowly resorbed, OP-1 enhanced mechanical stability of implants grafted with HA granules.

In study III we investigated the influence of OP-1 on the mechanical properties of bone allograft and HA-granules after three weeks. OP-1 increased all mechanical parameters of bone allograft and HA-granules.

In study IV and V we focused on platelet rich plasma (PRP) as a source of growth factor. We found no effect of PRP alone or in combination with bone allograft. We found no influence of processing by defatting, irradiation and freeze drying on the incorporation of impacted bone allograft.

Introduction

An increasing number of THA is done annually⁸¹. The 10 year survival is now reported up to 97 % and mean age at the primary replacement is 68 years⁴⁹. Therefore the majority of patients receiving their first replacement at an old age will never need a revision. Survival of an arthroplasty depends on age, gender and diagnosis and with a three years survival of 94% and 10 year survival down to 80% among men younger than 55 years, the number of revisions are expected to increase in the future^{49,81}. Aseptic loosening is estimated to be the cause of failure in 79% to 89% of all cases in Scandinavian countries^{49,115}. In general, revision THA have poorer clinical results, higher costs, longer recreation time and shorter longevity^{92,115}. One major problem in revisions is periprosthetic osteolysis and the success of revision arthroplasty is highly depended on restoration of the bone stock and creation of a stable implant.

The use of bone allograft (morsellized or/and structural) has become routine for many surgeons. Under optimal conditions the impacted morsellized bone graft provides initial mechanical support and creates a newly formed living bone stock which support the prosthesis and might make future re-revisions easier. *In vitro* studies show, that revision of the femoral component using morsellised bone graft followed by cementing with a collarless prosthesis with a polished tapered stem restores the integrity of the proximal femur and provides immediate stability of the implant⁸³. However histological analysis of retrieved impacted bone allograft often show incomplete bone incorporation^{17,79,93,161}. Follow up on cemented prosthesis inserted with impaction have not been conclusive. Reports from the innovators have shown good short- and middle term results at a level similar to primary arthroplasties⁸⁰. However high incidence of subsidence especially after massive loss of bone stock has been reported in later studies^{35,36,41,91,160} (table I).

A large number of bone growth factors has now been isolated and studied *in vitro* and *in vivo*. Among the BMP's, most focus has been put on BMP-2 and -7 which are now available for clinical use. The beneficial effect on fracture healing has now been documented in humans³⁹, but the effect on grafted and non-grafted implants is still unclear.

PRP is a concentrate of platelets in plasma containing a cascade of different growth factors and other cytokines capable of bone stimulation. By mixing bone allograft with purified growth factors or PRP bone invasion might be increased⁸⁸ and implant fixation and thereby possibly long term result could be improved.

Fresh frozen bone allograft for impaction is preferred in Denmark and other countries. The use of fresh frozen bone allograft is associated with possible immunological reactions, infections and limited supply however. Several processing techniques such as defatting, irradiation, treatment with antibiotics and freeze-drying have been suggested to decrease the potential disadvantages associated with the use of fresh frozen bone allograft²⁶. Bone substitutes are widely used in orthopaedic surgery, however no substitute has so far been used in larger scale to replace bone allograft in revision of failed THA¹⁰⁰. Ceramics such as HA or TCP might substitute bone allograft. However since the bone stimulating effect is mainly osteoconductive, a combination between a ceramic and a growth factor might be necessary to replace bone graft.

Aim (bliver udvidet)

The aims of this Ph.D. thesis is to answer the following questions:

- Can ProOsteon substitute or extend the volume of fresh frozen bone allograft?
- Is there any positive or negative effect of adding growth factor OP-1 to bone allograft or ProOsteon?

Stimulation and substitution of bone allograft around non-cemented implants

-Does PRP increase fixation and incorporation of implants grafted or non-grafted?
-How does processing of bone allograft effect implant fixation and bone incorporation? was to test the following hypothesis:

-In study I and II we tested the hypothesis that HA granules or a mixture of HA granules and bone graft can replace bone graft alone. Also we hypothesised, that the addition of OP-1 to the grafting materials increased the bioactivity of the graft and thereby increased stability of the implant.

-In study III we tested the hypothesis that OP-1 increases the mechanical properties of impacted HA granules whereas it impairs the mechanical properties of bone allograft due to accelerated bone graft resorption.

-In study IV we tested the hypothesis, that processing by defatting, freeze drying and irradiation impairs the bioactivity of fresh frozen bone allograft. Also we tested the hypothesis that PRP increases bioactivity of processed and fresh frozen bone allograft.

-In study V we tested the hypothesis that PRP increases implant bone incorporation and gap healing alone or in combination with bone allograft.

Background

Grafting in reconstructive hip surgery

Aseptic loosening of THA is often associated with lost bone stock on the femoral and acetabular site. Four different strategies can be used to obtain stability of the revised femoral stem.

- 1) A short stem designed to fill out the proximal osteolytic defect with proximal fixations is one possibility with minor bone loss.
- 2) Another approach is to use a long cementless stem with distal fixation leaving an empty proximal defect. This empty proximal defect may be left ungrafted.
- 3) Simple recementation with a long stem is a third possibility especially in older patients⁹².
- 4) A last possibility is reconstruction by impacted bone allograft around cemented or cementless stems.

Restoration of the bone defects by bone grafting may make future revisions easier. Therefore it is recommended to younger patients and to patients with major bone deficiencies.

Morsellized impacted bone grafting for cemented revision was introduced by Sloof in 1984 on the acetabular site based on Hastings and Parkes operation technique on protrusio acetabuli using autograft and cement¹³². On the femoral site, the use of impacted morselized bone allograft was first introduced without cement in Exeter in 1985⁴¹. Due to high subsidence, cemented technique was introduced in 1987 and subsequent development of operation technique and instrumentation was done as a cooperation between Exeter and Nijmegen. The first publications based on short term results were excellent. However, unpredictable early subsidence of the stem was later reported^{35,91}. At the beginning, impaction bone grafting mainly included less severe bone loss⁴¹, whereas later studies included more severe bone defects^{110,160} (table I). In one study, subsidence was found in 38% of the patients with an average subsidence of 10.1 mm⁹¹. High subsidence has been associated increased risk of loosening of primary inserted prosthesis⁵⁹ and also with pain⁶⁰. Recent studies have on revision THA have shown similar results when comparing cemented and cementless Exeter X-change technique¹¹⁰.

Stimulation and substitution of bone allograft around non-cemented implants

Subsidence and re-revision rates are dependent on multiple factors of which the individual surgeons experience and skills, prosthesis design and the size of the femoral defect seems to be the most important predictors^{94,110,160,162}.

Allograft is more often used in revision surgery than autograft due to required high volume of graft. Cortical or corticocancellous struts harvested from cadavers are mostly used to reconstruct uncontained bone defects. Special attention has been put on cortical struts to strengthen the cortical shell prior to impaction or cementation.

Table I: Results of femoral revisions using cemented impaction technique and collared stems

Author	Number hips Patients	Bone defect ^a	Follow up	Number of rerevisions	Subsidence
Gie, 1993 ⁴¹	56	Grade I: 9% Grade II: 48% Grade III: 43 Grade IV: 0	18-49 months	2	20% >5 mm 4% >10 mm
Elting 1995 ³⁶	56	Grade I 34% Grade II 40% Grade III 21% Grade IV 5%	2 years	2	48 % subsided
Eldridge ³⁵	79	No data	1 year	0	11% >10 mm
Melding ⁹¹	34	No data	30 months	2	35% 4-31 mm
Van Biezen ¹⁶⁰	21	Grade III: 43% Grade IV: 57%	41-85 months	0	5% >10 mm

a: bonedefects quantified according to Endo-Klinik

Risks and disadvantages associated with the use of bone graft

The use of bone allograft does carry potential risks of transmitting virus such as HIV, HBV and HCV^{2,10,86,95,124}. Even though the load of HIV in bone is low, experimental studies and clinical cases show, that HIV transmission is possible^{10,16,86,95,124}. One clinical case has been reported where bone graft was harvested from a cadaver in the U S. Fifty-three persons received bone grafts of whom four persons received fresh frozen bone allograft. Three of four patients receiving fresh frozen allograft were with HIV whereas none of the patients receiving freeze dried bone allograft were transmitted¹²⁴. Different processing techniques decreases the risk of transmitting virus. Ethanol destroys HIV. However, there is concerns if ethanol penetrates cortical bone sufficiently. Bone marrow has a heavy load of HIV which is decreased by defatting and lavage¹⁰. Today's practise is screening the donor for hepatitis and HIV and no cases of HIV transmission have so far been reported using graft from tested donors.

Bone grafts also carry the risk of bacterial contamination. Even though cultures taken at the time of harvesting are negative, they might occasionally be positive when it is later thawed and result in

deep infection^{2,141,146}. Again, processing of the allograft by soaking in antibiotics, ethanol or irradiation decreases the risk of contamination. However, even though various methods of preserving, processing and sterilization might decrease the risk of transmission, the most important prevention is sufficient medical and social history of the donor and donor screening. Even though certain donors are excluded, the mineral density of the donated femoral head varies with the general bone quality and hence the quality of banked bone allograft is not consistent^{47,109}. The demand of bone allograft depends on the number of revisions and also the number of femoral heads needed in each operation. Both parameters are expected to rise⁴⁰ and the supply of femoral heads might not be sufficient in the future. Consequently alternative operation techniques to impaction such as distal fixation or substitutes to bone allograft might be used in the future.

Biomechanics of impacted morselized bone allograft

The biomechanical properties of morselized bone allograft alone have been investigated in different experimental set-ups^{14,42,155}. Impacted morselized bone allograft has the mechanical characteristics of an elasto-viscoplastic material⁴². The impacted bone recoils immediately after impaction which explains the relatively high initial stability of the prosthesis. Stability of a grafted implant is dependent on a number of parameters such as impaction pressure, chip size and bone graft properties and type of prostheses^{153,155}.

Freeze-drying of trabecular bone increases strength and stiffness, however rehydration makes freeze-dried bone similar to fresh frozen bone allograft¹⁹. The combination of freeze-drying and irradiation seems to weaken the trabecular bone²⁴ and freeze-drying prior to irradiation is worse than the irradiation prior to freeze-drying¹¹⁴.

Bigger size and partly defatted bone chips increase stability of cemented cups¹⁵³. Usually bone chips with diameters from 3-5 mm are recommended. Also higher impaction force increases initial fixation however cautions should be taken to prevent fractures⁹¹. In cadavers, the initial stability of cemented stems inserted with impacted bone allograft is lower than cemented stems inserted without impacted graft but better than uncemented stems with no graft^{11,83}.

The biomechanical properties of impacted bone graft changes when tissue invades the graft. Fibrous ingrowth alone increases strength¹⁴⁸. The biomechanical properties of bone chips impacted into rabbit condyles were similar to the original bone after three weeks¹⁷³. However the bone ingrowth was better than would be expected in bone allograft in humans.

Incorporation of impacted bone allografts

Lamerigt described the events of bone incorporation of impacted bone allograft as *"invasion of the bone graft by a front of vascular fibrous tissue after which osteoclasts resorbed the dead bone graft, followed by woven bone apposition on the graft remnants"*⁷⁰. This front does not go all the way from the periphery to the cement or implant surface. Usually three layers in the grafted area around cemented implants can be identified histologically: an inner zone with fibrous tissue and pieces of allograft chips, a middle zone with bone formation and neocortex and an outer zone consisting of cortex⁹³. Bone incorporation of impacted bone allograft is fastest during the first six months⁷⁹ and radiographic examinations show little change after two years³⁶.

Specimens of impacted graft have been taken as biopsies or autopsies in humans around cemented acetabular and femoral components (table II). Usually a higher degree of bone ingrowth into the graft is seen on the acetabular side compared to the femoral site. Incorporation of impacted morselized bone allograft has also been tested in various animal models. However since bone turnover is higher in the usually young animals and because the anatomy often allows only smaller gaps, bone incorporation and graft remodelling are often reported to be greater than the findings from human studies^{73,126,173}.

Stimulation and substitution of bone allograft around non-cemented implants

Many factors influence the rate and extent of bone graft incorporation including immunological response, processing of the graft, size of the bone chips and porosity of the impacted graft particles, thickness and tightness of the graft, load and stability and the bone forming capacity of the recipient.

Table II: Histological findings in incorporated impacted bone allograft in humans around implants

Implant	Source of specimen	Number of specimen	Observation time	Ingrowth	Graft	Author
Cemented femur Component	Autopsy	1	6 months	Fibrous invasion in the proximal part	Graft mostly embedded in fibrous tissue	Ullmark ¹⁵⁴
Cemented femoral component	Biopsies	4	11-27 months	Fibrous tissue in the innerzone, new bone in the periphery	Bone chips embedded in fibrous tissue	Nelissen ⁹³
Cemented femoral component	Biopsies and autopsies	14	3-96 months	Usually fibrous invasion	Still chips after 8 years	Linder ⁷⁹
Acetabular Component	Biopsies	8	1-72 months	Fibrous tissue dominated in the innerzone, bone in the periphery	Bone chips remodelled in the periphery, few chips in the innerzone	Buma ¹⁷
Acetabular component	Biopsies	24	3-180 months		Non-incorporated graft in some areas regardless of time	Van der Donk ¹⁶¹

Allografts are rarely tested for histocompatibility even though frozen bone allograft contains cell debris with antigens. Studies have shown, that recipients are being sensitised towards the HLA-type of the graft¹⁴⁴. Mismatched fresh bone allografts are poorly integrated^{56,144} and host immune response might play a role in incorporation^{43,44}. One approach to impair the rejection could be systemic treatment with immunosuppressive drugs used for transplantation of organs^{43,44}. However taken the side effects into account, that is not clinical relevant. Another approach to decrease the immunological response is to decrease the load of antigens. Freezing alone decreases the immunological response¹⁴⁴. Removing the bone marrow cells by lavage and defatting improve bone incorporation^{9,56,151} probably because the load of cells is decreased. Removal of cartilage from the femoral head is of great importance of bone incorporation¹⁶¹.

The impact of load on incorporation of bone graft has been proved in a number of experimental studies¹⁶³. Wang showed in a rabbit tibia that a loaded stem increased graft resorption and bone formation compared to an unloaded stem¹⁶⁷. Moreover Donk S found a bigger area of active bone incorporation in loaded impacted bone compared to unloaded¹⁶³. Since bone around a femoral component is not equally loaded, the histological findings of graft around prostheses might differ according to the anatomical location with more graft replaced by bone distally compared to more proximal¹⁵⁶.

Stimulation and substitution of bone allograft around non-cemented implants

The speed of bone formation and the mechanical properties during remodelling differs between cortical and trabecular bone due to differences in porosity. Massive cortical grafts are gradually resorbed prior to invasion of vessels and might only be partly substituted by bone^{37,46}. The fact that cortical graft is resorbed prior to new bone formation might lead to mechanical failure even though it is initially mechanical sufficient³⁷. On the contrary, cancellous bone allograft serves as a scaffold, where bone is formed on the surface of the trabecular of the bone graft. For that reason, mechanical strength of cancellous graft tends to increase during invasion of bone. The bone allograft is gradually resorped during remodelling of the construct. By soaking the graft in a solution of bisphosphonates, resorption of the graft can be postponed⁷.

Porosity is decreased by impaction of the morsellized bone allograft, compared to cancellous bone¹⁴⁷ delaying bone incorporation. Whether the mechanical properties of impacted bone allograft during remodelling is similar to cortical or cancellous bone is not well known.

The importance of BMP's in non-demineralized bone allograft is still not clear. Processing of bone allograft chemically, by irradiation or heat might inactivate BMP's and other growth factors contained in the bone matrix. The growth factors can be demasked by demineralising the bone (DMB). Such bone graft is commercial available and has been proven to be osteoinductive and will usually be incorporated with bone faster than non-demineralised allograft. Application of DMB around implants has so far not been encouraging²².

Bone graft substitutes

A number of bone graft substitutes has been suggested to replace morsellized bone allograft around implants due to the disadvantages associated with bone allograft described above. Although e.g. growth factors and bone marrow aspirates improves gap healing, they do not provide initial mechanical support and can therefore not replace bone allograft in revisions. Neither can DBM due to the poor mechanical properties.

Mechanical properties of ceramics like HA and TCP granules have been tested^{12,152,164,173}. They do not have the visco-elastic properties as previously described for bone allograft. Therefore they will not recoil and the initial fixation of implants surrounded by only HA or TCP granules might not be sufficient¹⁶⁴. However stability of cemented femoral prostheses grafted with porous HA/TCP composite and bone allograft have shown promising results^{12,164}.

Table III: Animal studies on bone graft and bone graft substitutes around non-cemented implants

Graft	Model	Observation periode	Animal	Resultat	Author
Autograft	THA-revisions, fiber-Ti implants	12 weeks	Canine	+++	McDonald ⁹ ₀
Allograft	Revision THA, fiber Ti implants	12 weeks	Canine	+++	McDonald ⁹ ₀
Autograft	Revision THA, fiber Ti implants	6 months	Canine	+++	Turner ¹⁵²
Autograft	Revision THA, fiber Ti implants	6 months	Canine	+++	Turner ¹⁵²
HA/TCP granules	Revision THA, fiber metal Ti implants	6 months	Canine	0	Turner ¹⁵²
Allograft	2 mm gap model, Ti implants	6 weeks	Canine	+++	Soballe, K ¹³⁹
Allograft	2 mm gap model, HA-coatede implants	6 weeks	Canine	+++	Soballe, K ¹³⁹
Allograft	THA, HA-coated implants	6 og 12 weeks.	Goat	++	Schreurs, B W ¹²⁶
HA-granules	3 mm gap model, HA-coated implants	3 weeks	Canine	0	Study I
Allograft/HA	3 mm gap, HA-coated implants	3 weeks	Canine	++	Study II

Stimulation and substitution of bone allograft around non-cemented implants

granulae					
Allograft	2.5 mm gap model, HA-coated implants	3 weeks	Canine	+++	Study IV

Ceramics have not shown same level of bioactivity as bone allograft^{27,152} but one recent study has indicated, that ceramics might be used as an extender of bone allograft around femoral stems¹¹¹. Resorption of ceramics depends on structure and chemical compound. TCP is resorbed faster than HA^{15,31,129}. Due to the differences in mechanical properties between ceramics and bone graft and since some HA granules might never resorb, bone incorporated HA granules might never obtain the same mechanical characteristics as bone alone¹⁷³. It is not known if implants surrounded by such composite have inferior survival.

Also Bioglass (45S5) has been suggested to replace bone allograft. Bioglass is incorporated at a faster speed compared to HA granules and is faster resorbed^{99,101}. Bioglass has recently shown promising results as an extender of bone allograft around revision hip prosthesis³⁴.

Bone growth factors

Bone growth factors are polypeptides secreted by bone- and other cells (table IV) providing a mechanism for altering cell behaviour such as division, differentiation and matrix synthesis. The exploration and identification of bone growth factors have taken more than a century. In 1889 Nicholas Senn used bovine demineralized bone matrix to fill defects after osteomyelitis and found, that new bone developed. Huggins discovered in 1931, that epithelia from bladder and urether resulted in ectopic bone formation when it was implanted in fascies of guinea pigs. A mile stoe was the article in 1965 by Urist discovering that DBM induced ectopic bone bone formation and later he isolated BMP's which had greater bone inducing effect than DBM¹⁵⁷⁻¹⁵⁹. BMP's were initially isolated from bovine bone until 1988 when Wosney produced recombinant human BMP-2¹⁷¹.

Bone growth factors have a number of potential surgical applications might be used in the future in a number of clinical situations including treatment of pseudoarthrosis secondary to impaired bone healing, spinal fusions and bone reconstruction after loosened implants and tumor resection.

Table IV: Examples of growth factor sources

Growth factor	Source
TGF-β	Platelets, leucocytes, osteoblasts, chondrocytes
BMP	Chondrocytes, osteoblasts, urether- and bladder epithel
FGF	Monocytes, macrophages, osteoblasts, chondrocytes
PDGF	Platelets, monocytes, macrophages

At the moment bone growth factors BMP-2 and BMP-7 are commercially available. OP-1 (BMP-7) is approved in the treatment of nonunion of the tibia of at least 9 month duration, secondary to trauma in skeletally mature patients, in cases where previous treatment with autograft has failed or use of autograft is infeasible³⁹. Also DBM and kits for preparing PRP are commercialiced.

Platelet concentrates

Fracture repair is regulated by a number of systemic and local factors. In all stages of fracture repair, growth factors, cytokines and other proteins produced by platelets, leucocytes, and macrophages are believed to play an important role^{88,143}. A haematoma is formed at the site of a fracture; platelets are activated by collagen exposure leading to fibrin clotting and platelet

aggregation. Thrombin initiates platelet degranulation. A large number of growth factors such as platelet derived growth factors (PDGF), transforming growth factor- β (TGF- β 1), insulin-like growth factors (IGF) and epidermal growth factor (EGF) have been isolated from platelet α -granules^{71,71,71,131,131,131,169}. Other factors such as β -thromboglobulin and macrophage inflammatory protein-1 α , which are mediators of inflammation with various effects on white cells including macrophages are also found in platelet granules⁶⁵.

Platelet concentrates are usually used in a fraction of autologous plasma. Marx introduced the name "platelet rich plasma". Later a number of different companies has patented kits for PRP preparation and call the final product AGF, Symphony, PCCS etc.

PRP is basically based on the idea that "more is better". Local application of PRP might increase the local concentration of growth factors and thereby stimulate healing of damaged tissue such as bone defects or ulcers^{66,67,84}. Since platelets are active in fracture and tissue repair, a concentrate might be beneficial. Furthermore PC stimulates osteoblast proliferation *in vitro*^{131,131,131}. Local application of platelet concentrates (PC) has been suggested to enhance healing of damaged tissue such as bone defects or ulcers{24}{464}{465}.

Experimentally, platelet concentrate increased bone ingrowth in bovine cancellous bone in a rabbit calvarial defect model⁶². Also Kim et al found more bone in contact with dental implants in the group treated with DBM and PRP than DBM alone after 6 weeks but not after 12 weeks⁶³. Jensen et al found effect of AGF®, a platelet concentrate processed with the use of a commercialized filter, in combination with allograft around non-HA coated implants in canines⁵⁴. However they found no effect of AGF® with no bone graft around loaded implants⁵⁴ indicating, that platelet concentrates need to be mixed with a material to keep it on location preventing it from dilution.

Platelet concentrates in a number of articles describes the use of platelet rich plasma to enhance bone regeneration and healing of soft tissue in humans^{4,66,142,165,170} but only trial using PRP to enhance bone healing has been published⁸⁸. In a prospective randomized clinical study, 88 patients with mandibular defects were randomly treated with morselized autograft alone or autograft+PRP. PRP was increased by 238% compared to base line. After 6 months more bone was found in defects treated with PRP and radiographic examination showed enhanced maturity and bone consolidation of the PRP treated graft.

A number of companies has recently commercialized kits to prepare PC in autologous plasma. The level of platelets and growthfactors also level of fibrinogen might depend on the commercial kit being used. One important question to answer regarding the use of PRP is the optimal concentration of platelets. It is well known, that growth factors only stimulate bone healing beyond a certain threshold. Since most growth factors are found in bloodcells and not in the plasma, a correlation between level of bloodcells and bone healing is expected. However this has only been proved on TGF- β 1¹⁶⁹.²⁹ Since growth factor levels might be individually associated with sex, age, smoking etc., the growth factor levels should ideally be determined in each preparation to ensure activity. But since PRP preparation is often done at the same time as the operation takes place, platelet counts as a quality control is considered good manufacture practise²⁹.⁴

By adding thrombin to the plasma, a gel is formed and platelets are activated¹⁷⁵. Theoretically not only platelets but also the gel might stimulate bone formation. Fibrin adhesives have been used in many surgical areas since early 70's. It can be prepared from autologous¹⁴⁹, single donor or pooled plasma samples. Fibrinogen products such as Tisseal® (Immuno, Austria) are commercially available and is used for numerous applications to seal lungs, trachea, vascular anastomoses and liver- and kidney injuries. Fibrin sealant makes bone chips easier to handle^{88,149}, however the influence on bone healing is controversial⁸². Bone might be formed on the surface of fibrin as reported from biopsies from impacted bone allograft¹⁶¹.

Stimulation and substitution of bone allograft around non-cemented implants

Table V: Platelet concentrations in a number of platelet concentrate studies

Preparation	Animal	PRP Platelet conc. (*10 ⁹ /l)	Mean increase (%)	Reference
No commercial kit	Human	785	383	Marx RE ⁸⁸
No commercial kit	Human	3990	1700	Dugrillon, A ²⁹
PCCS®	Human	2209	761	Weibrich, G ^{168,169}
Curasan	Human	1075	371	Weibrich, G ^{168,169}
No commercial kit	Canine	1735	391	Kim, ES ⁶¹
AGF®	Canine	1212	717	Jensen, TB ⁵²
PCCS®	Canine	645	496	Jensen, TB ⁵⁴
No commercial kit	Canine	1884	770	study IV and V

Bone growth factors and grafted implants

A large number of purified growth factors has previously been investigated in combination with bone graft and bone substitutes in bone defects. BMP-3 stimulates bone ingrowth in HA in baboons ^{116,117}. BMP-2 alone increases ingrowth in HA in calvaria defects in rabbits ^{96,97} and OP-1 has previously increased ingrowth in HA in sheep ²⁷. bFGF stimulates bone ingrowth and increases vascular invasion into bone grafts and porous HA depending on dose ^{69,166}. Addition of rhBMP-2 to autograft increases the expression of other BMPs and genes associated with bone formation and is capable of increasing bone formation in the central zone of spinal fusion masses in rabbits. In an unloaded bone chamber, the addition of rhBMP-2 increased the number of osteoclasts and amount of woven bone and fibrous tissue.

Since it is well documented, that growth factors increase bone incorporation of bone graft and HA, growth factors might as well increase bone incorporation of grafted titanium implants. Soballe et al, found no effect of OP-1 on fixation of loaded implants in a primary setting. But in a revision with delayed bone healing properties, OP-1 increased fixation, however only at low concentrations ¹³³. This indicates, that the greatest response of growth factors might be found in models and clinical situations with impaired healing capacity. Also OP-1 has been used in a clinical trial in combination with bone allograft in the revision of failed THA. However the trial was stopped due to some early failures ⁶.

Table VI: Enhancement of bone allograft implants using growth factors and platelet concentrates

Stimulation and substitution of bone allograft around non-cemented implants

Growth factor	Carrier	Graft	Model	Obs. period	Animal	Result	Ref.
rhOP-1	No carrier	fresh frozen bone allograft	loaded, non-HA coated, gap, trabecular bone	4 weeks	canine	no effect on fixation	Søballe, K ¹³³
rhOP-1	No carrier	fresh frozen bone allograft	revision, loaded, non-HA coated, gap, trabecular bone	4 weeks	canine	effect at lower concentration	Søballe, K ¹³³
AGF®	No carrier	fresh frozen bone allograft	unloaded, non-coated, gap, trabecular bone	3 weeks	canine	effect on bone ingrowth and fixation	Jensen, TB ⁵⁴
PRP	No carrier	fresh frozen bone allograft	unloaded, HA coated, gap, trabecular	3 weeks	canine	no effect on bone ingrowth or implant fixation	Study IV
PRP	No carrier	fresh frozen or processed bone allograft	unloaded, HA coated, gap, trabecular bone	3 weeks	canine	no effect on bone ingrowth or fixation	Study V
rhOP-1	Collagen	fresh frozen bone allograft	unloaded, HA coated, gap, trabecular	3 weeks	canine	effect on bone ingrowth	Study I
rhOP-1	Collagen	ProOsteon	unloaded, HA coated, gap, trabecular	3 weeks	canine	effect on bone ingrowth and implant fixation	Study I
rhOP-1	Collagen	ProOsteon/fresh frozen bone allograft	unloaded, HA coated, gap, trabecular	3 weeks	canine	effect on bone ingrowth and implant fixation	Study II
rhOP-1	Collagen	Bone allograft	loaded knee prostheses	6 weeks	rabbit	no effect	Jeppsson, C ⁵⁵
rhOP-1	Collagen	Fresh frozen bone allograft	unloaded, HA coated, gap, trabecular	6 weeks	canine	no effect	Lind, M ⁷³
PCCS®	No carrier	Allograft	Non-loaded, non-HA coated, gap, trabecular bone	4 weeks	canine	effect on bone ongrowth and fixation	Jensen, TB ⁵²

Bone growth factors and non-grafted implants

The ability of increasing bone incorporation of ungrafted implants has been investigated in a number of studies. TGF- β absorbed to TCP or HA coatings increases implant fixation and gap healing, however the effect is highest at lower concentrations^{25,75,76,140}. Cook et al found no effect of OP-1 on bone ingrowth in dental implants inserted press fit after 12 weeks²³. OP-1 in a collagen carrier had little effect on implants inserted in trabecular bone surrounded by a gap.

Table VI: Studies on enhancement of implant fixation using growth factors or PRP with no graft

Growth factor	Carrier	Model	Obs. period	Animal	Result	Ref.
Bovine OP*	Collagen	Interference fit after tooth extraction, non-HA coated	3 weeks	Monkey	Qualitative evaluation, increased ingrowth	Rutherford, RB ¹²¹
rhOP-1	Collagen	Unloaded, +/- HA coating, gap, trabecular bone	6 weeks	Canine	Effect on fixation bone ingrowth and gap healing compared to empty gap but only on fixation compared to collagen	Lind, M ⁷⁸
rhOP-1	Collagen	Interference fit after tooth extraction, +/- HA coating	12 weeks	Canine	No effect	Cook, SD ²³
TGF- β 1	Implant	Loaded +/- TCP coating, gap	6 weeks	Canine	Effect on ingrowth, no effect on fixation	Lind, M ⁷⁵
TGF- β 1	Implant	Unloaded +/- TCP coating, gap	6 weeks	Canine	Effect on fixation and bone ingrowth	Lind, M ⁷⁶
TGF- β 1	Implant	Unloaded, HA coated, gap	6 weeks	Canine	Effect on fixation and bone TGF- β 1	Lind, M ⁷⁴
TGF- β 1	Implant	Unloaded, HA and TCP coated, gap,	4 weeks	Canine	Effect on bone ingrowth and gap healing	Sumner, RE ¹⁴⁵
PRP	No carrier	Unloaded, HA coated, gap, trabecular bone	3 weeks	Canine	No effect	Study IV and V
AGF	No carrier	Loaded, non-HA coated, gap, trabecular bone	4 weeks	Canine	No effect	Jensen, TB ⁵²

*bovine OP consist of BMP-2A and OP-1¹²⁵

Materials and methodological considerations

Ethical considerations

Canines bred for research were used in all studies. All animal handling was approved by Danish Control Board for Animal Research.

Animals

A long list of animals from mice to baboons are being used in orthopaedic research. We chose canines for several reasons. Observation time, animal mobilisation and post operative care depend on species and we have more than 10 years of experience using canines allowing us to compare new data with previous results. Also canine bone has similar composition as human bone¹. The size of the animal is critical using the Soballe model and the use of smaller animals such as rabbits is not possible. Animals such as goats or sheep have been used in implant research and might be used in our research group in future projects³.

We used same breed of dogs, at approx. same age and weight to decrease the biological variation.

Observation time

We used a relatively short observation period of three weeks for several reasons. Remodelling in canine bone has been estimated to be 2-3 times that of humans⁶⁴. We used young, but skeletal mature, healthy dogs and did not do any attempts to decrease bone healing.

A gap of 2.5 mm surrounding a HA coated implant inserted in trabecular bone in a healthy canine heals after 6 weeks¹³⁹. Therefore all the gaps in our studies, grafted or non grafted, would probably heal after a time period and a short observation time was thus essential. A short observation time is interesting from a clinical point of view since early anchorage of the implant is an important predictor of long term survival⁵⁹.

Implants

We used porous coated titanium implants plasma sprayed with HA. Titanium porous surfaces are osteoconductive and the bioactivity is further enhanced by HA coating¹⁰². As a consequence Soballe et al found strong effect of bone grafting in gaps surrounding non-HA coated implants but no effect of bone grafting when the implants were HA coated¹³⁹. Differences between treatment groups might easier have been demonstrated using non-HA coated implants. HA coatings similar to the one we used in our experimental studies now show good clinical results^{85,119}.

Experimental model

Implants were surrounded by 3.0 mm gaps (study I and II and III) or 2.5 mm gaps (study IV and V). The implants were non-loaded and stable inserted extraarticularly in the femoral condyles¹³⁹ (study I, II and III) or the proximal humerus¹⁰⁵ (IV and V).

This model was developed by Soballe and has been used for more than a decade in Orthopaedic Research Group to investigate the influence of various surface coatings and textures, growth factors, bone allografts, operation techniques and bone substitutes^{53,74-78,102-105,107,112,113}.

The border of the drill hole was surrounded by trabecular bone in the femoral condyles whereas the border of the drill hole in some areas was in contact with cortical bone in the humerus. Primary cementless THA rely on cancellous bone ingrowth in the proximal femur. However in revisions

with extensive bone resorption, only a cortical shell might be left or the defect might even be uncontained.

We used a non loaded model allowing no micromotions to occur. Implants in the clinical settings are loaded eventhough full weight loading is often avoided the first months postoperatively. Load plays a keyrole in incorporation and remodelling of bone allograft (Wollfs law). This model is thus less clinical relevant compared to other models previously been used in experimental orthopaedic research. The main reason for choosing the non-weight loaded model was, that it has less “noise”. We could have used a loaded Soballe model^{103,104,107,112,113,135-138}. However this model only allows a gap of 0.75 mm which might be too small to further increase bone grafted implants by adding growth factors.

THA has previously been inserted in canines and goats¹²⁶⁻¹²⁸. This provide more clinical relevant data. However important parameters such as the volume of the graft, load and micromotions might not be easy to standardize in such model so the biological variation might increase significantly. Therefore more animals would be needed and only one grafting material would be investigated in each animal increasing the expenses and number of animals.

Design

The size and anatomy of the canines allowed insertion of implants in trabecular bone in each femoral condyle and two implants in each proximal humerus. Two studies were designed in each dog, each study using a paired design. Due to differences in loading and possible differences in mineral densities between humerus and femur, data humerus and femur should not be compared. Also, bone healing in the medial vs the lateral condyle in the femur and proximal vs distal humerus might differ due to differences in bone density. The different treatment groups were block randomized to the different locations. The paired design within each animal gives a higher statistical power since variation is decreased⁶⁸.

Sample size

The error of first kind (2α) was selected to be 5% and error of second error (β) to be 20%. Based on previous studies we estimated SD to be 50% of the mean. The minimal difference between treatment groups to be detected was set to be 70%. Based on these assumptions, the minimum number of animals was calculated to be seven¹⁰². Eight dogs was thus included in each study.

Grafting materials

Fresh frozen bone allograft (study I-V)

Proximal humerus, proximal femur and proximal tibia were harvested from two dogs in study I-II and III and one dog in study IV and V. After three weeks the bone graft was thawed and soft tissue and cartilage was removed. Using the finest grater in a standard bone mill, the graft was milled to chips which could be used in a 2.5 mm gap. These chips sizes are smaller than recommended in revision surgery. Smaller chips incorporate and resorb faster¹⁰⁸. Bacterial cultures ensured, that the graft was not contaminated. The graft was packed as tightly as possible. Impaction does decrease ingrowth, however tight impaction ensures implant stability.

Processed bone allograft (study IV)

Freshly morsellized bone allograft was further processed by lavage, defatting in ethanol, freeze drying and irradiation. These steps are similar to recommendations by American Red Cross.

ProOsteon (study I,II and III)

ProOsteon 200 (Interpore,Irvine,US) is a corraline porous hydroxyapatite bone substitute approved by the FDA which has been on the market for more than two decades. The osteoconductive properties are well documented. Coral originally consists of 99% calcium carbonate¹²⁰ which is chemically transformed into almost non-resorbable HA¹²⁰. ProOsteon 200 was delivered as granules with a diameter of 425 µm to 1000 µm and a mean porous diameter of 230 µm and interconnection diameter of . Void fraction of ProOsteon 200 is 63%, mean trabecular thickness is ProOsteon was standardized by weight, put into containers and autoclaved according to manufactures instructions.

OP-1 device (study I,II and III)

OP-1 was used in study I, II and III. OP-1 is considered one of the most potent bone stimulating growth factors. Furthermore, it is commercially available for clinical use.

Recent studies on OP-1 have not encouraged further use of OP-1 in revision surgery, however at the time of our experiment no studies on the effect of OP-1 on bone allograft was published. We could as well have used BMP-2 which was available through another company.

OP-1 was delivered in a device consisting of 2.5 mg recombinant human OP-1 in 1 gram of bovine type I collagen (Stryker Biotech). Using this formula, we can not conclude whether the effect of OP-1 is due to OP-1 or the collagen carrier. Bovine collagen type I stimulates human osteoblasts *in vitro* and collagen enhances integration of bone substitutes *in vivo*. Collagen has osteoconductive properties with high bioactivity probably because it is capable of binding circulating factors such as osteonectin and growth factors. Surprisingly, collagen I alone proved to be just as efficient as collagen+OP-1 to promote bone ongrowth to non-cemented HA coated and non-coated implants emphasizing the high bioactivity of collagen.

The effect of OP-1 depends on concentration^{20,21}. The dosage of OP-1 in the present study was 300 µg OP-1 in 120 mg collagen carrier used in a 0.75 cc gap. Determination of concentration was based on studies by Cook in non-unions in canines^{20,21}. He concluded that concentrations beyond a certain threshold did not further increase bone formation.

Platelet Rich Plasma (study IV and V)

The preparation of PRP was done following the same procedure as described by Marx⁸⁸.

Platelets and leucocytes counts in the PRP were appr. 770% and 910% compared to venous blood (table VIII). Also erythrocyt count was increased.

Table VIII, analyses of PRP and whole blood. PRP/whole blood was calculated for every single dog. (median(range))

Group	Baseline Count	PRP	PRP/whole blood
Platelets (*10 ⁹ /l) n=8	246 (132-321)	1884 (1156-2742)	7.7 (6.0-8.9)
Leukocytes(*10 ⁹ /l) n=8	8.1 (5.8-13.3)	71.7 (45.1-95.5)	9.1 (6.8-11.8)
Erythrocytes(*10 ¹² /l) n=8	6.0 (5.0-7.3)	8.6 (7.6-11.6)	1.5 (1.2-2.0)

Given our negative results in study IV and V, a test of the level of growth factors or a positive test on a cell culture would have been recommended.

At the time of surgery, the commercial kits for preparation of PRP such as Symphony and AGF were not available.

Mechanical evaluation

The object of the mechanical test in study I, II, IV and V was to evaluate the bone-implant surface interface mechanically. The fixation of an implant is determined by the direct bonding of tissue, of which bone is believed to give the best fixation. Also bone interlock on a porous coated implant might play a role¹⁰². We used a destructive push-out test which has been used in several studies. A pull-out or torque test could have been an alternative. However no mechanical test can mimic the clinical load, which is not only axial but also involves bending, shearing and compression. As an alternative to the destructive push-out test, we could have chosen a cyclic test⁵¹.

Thickness of implants varied from 2.8-3.6 mm and push-out data was normalized by the surface area of the tested implant. Clearance (distance from the surface of the hole in the support jig to the surface of the implant) was set to 500 µm as suggested by Dhert²⁸.

Ultimate shear strength, stiffness and energy absorption were determined on the load-displacement curves as previously described^{102,134}. Energy absorption has been suggested to be the most important mechanical parameter¹⁰². A high stiffness might minimize micromotions, but a low energy absorption could lead to failure¹⁰².

The outcome of push-out test is affected by a number of parameters¹⁰². We reduced the potential risks of variations due to storing, machine calibration, temperature, centralization over the support jig etc. by doing the tests in each study the same day, in random order and done blindly.

The aim of the mechanical test in study III was to test the mechanical properties of incorporated bone graft and ProOsteon. This test was performed by centralizing the grafted 11 mm gap over a hole of 11.3 mm in diameter. A piston 10 mm in diameter applied load on the gap (paper III, figure 1 and 2). This test is a combination of compression of the bone loaded by the piston and the metal platform and a push-out test with stress applied to the interface at the border of the drillhole. The failure was always seen at the border of the drillhole. A similar test has previously been used to test newly formed bone in craniotomies¹⁸.

Histomorphometry

After dehydration, each specimen was embedded in methylmethacrylate (Technovit 7200 VLC, Exakt, Germany). Four sections of 25-30 µm thickness were cut on a microtome (Leiden, Holland) and surface stained with 2 % light green⁴⁵. By this staining method, mineralized tissue is stained green other tissue is red.

In study I and II, the sections were done perpendicular to the long axis of the implant. In study III and IV, the vertical section method¹⁰⁶ was followed: Each implant was randomly rotated around a vertical axis of the implant prior to sectioning and serial sections were made parallel to that axis. Quantification was performed using an image-analysis system (Grid, Olympus, Denmark). The microscope fields were transmitted to a computer screen and user-specified grids were superimposed randomly according to the method for unbiased estimates¹⁰⁶. The vertical method gives an unbiased estimate surfaces. However volume fractions can be estimated unbiased using either methods. Using the vertical section method will only present a true value of the size of the peri-implant gap when the section is done through the centre of the implant¹⁰².

Volume fractions of woven bone, grafting material and other tissue in the gaps were determined in two well defined zones: Respectively 0-1 mm from the implant surface and 0-1 mm from the border of the drill hole at a 100X magnification. 250 points were counted in each of the two zones bone on every section. In order to estimate bone coverage of the implant, 250 intersections between a line grid and the surface of the implant was counted on each section.

The influence of OP-1 on the density of the bone surrounding the drill hole was studied in study I. 420 points were counted in a 1 mm zone outside the border of the drill-hole (zone 3) and volume fraction of bone was determined.

In order to compare volume fractions of grafting materials in study one and two after 3 weeks to those at the time of implantation, eight control implants from each treatment group were inserted into cadaver bone using the same materials as in the *in vivo* experiment. The control implants and surrounding bone were cut out *en bloc* and prepared and evaluated as described previously.

Reproducibility

Double measurements on histomorphometry on all sections from the ProOsteon and allografted group (a total of 12 implants) were done in study II with a time interval of approximately 2 years by the same person. Reproducibility (intra-observer variation) was calculated as coefficient of variation (CV) as previously described¹⁷⁴:

$$S^2 = (1/2k) \sum d^2,$$

Where k is the number of double measurements (in the present study 6 in each group) and d is the difference between first and second quantification. CV is calculated as

$$CV = s/x$$

Where x is the mean value of first and second quantification.

CV was highest on bone ingrowth. CV on bone graft was higher than CV on ProOsteon (table IX).

Table IX, study II: Coefficient of variation (CV) based on double measurements in percentage

	Bone ingrowth	Woven bone		Soft tissue		Graft/ProOsteon	
		Zone 1	Zone 2	Zone 1	Zone 2	Zone 1	Zone 2
Allograft, n=6	6	9	5	3	4	7	12
ProOsteon, n=6	9	6	4	2	5	5	6

Statistics

Most of the data was not normal distributed and non-parametric tests were chosen in all studies. one way ANOVA on ranks was applied to determine any significant differences between four groups. Groups were pairwise compared using Student-Newman-Keul or Wilcoxon signed rank test.

Data are presented as median (range) in study I, II, III and V and median (interquartile range) in study IV. P values less than 0.05 were considered significant.

Results

Exclusions

Two of eight dogs were killed after two weeks in study I, II and III (same dogs were used in all three studies). In study IV, four implants were excluded prior to evaluation since they were inserted too close to the knee joint cavity. No samples were lost during preparation or analysis.

Table X, study IV: Numbers of dogs in each treatment group after exclusion of displaced implants

Group	Fresh frozen allograft	Processed allograft
Without PRP	n=6	n=7
With PRP	n=7	n=8

Study I

Mechanical test

Bone allograft had significantly better fixation compared to ProOsteon without OP-1 device. Adding OP-1 device to bone allograft resulted in an insignificant decrease of ultimate shear strength and stiffness. In contrast, OP-1 device increased fixation in the ProOsteon group. OP-1 device increased ultimate shear strength of ProOsteon by 800% to a level comparable with bone allograft (table XI).

Table XI, study I: Push-out values median (range)

Group	Ultimate shear strength (MPa)	Energy abs. (J/m ²)	Apparent Stiffness (MPa/ mm)
Allograft	2.30 (0.49-4.75)	330 (144-719)	16.1 (13.0-26.0)
ProOsteon	0.39 (0-0.58) a	83 (0-357) a	1.4 (0-2.1) a
Allograft+OP-1 device	1.91 (0.38-4.83)	346 (61-705)	13.1 (7.5-35.9)
ProOsteon+OP-1 device	2.56 (0.82-5.67)	382 (205-719)	25.9 (4.3-50.3)

a: p<0.05 compared to the three other groups<

Histology

Qualitative analysis: New bone formation was deposited mainly on the surface of bone allograft (paper I, figure 3a and b) or ProOsteon granules (figure 4a and b). In the ProOsteon group, bone apposition was seen on the HA coating despite absence of bone formation in the gap close to the implant. Remnants of OP-1 collagen carrier were found in a few gaps. Resorption lacunae could be

Stimulation and substitution of bone allograft around non-cemented implants

recognised on the surface of the allograft and ProOsteon. Non-mineralised tissue was mainly soft and cell-rich.

Quantitative analysis: No significant differences in bone ongrowth to the implant between the four groups were found. OP-1 device significantly increased bone formation in the gap in both zones and in combination with both grafting materials (table XII). In zone 1, 6 (0-13)% woven bone was seen in ProOsteon without OP-1 compared to 12 (7-20)% in bone allograft without OP-1. In zone 2, 23 (15-26)% woven bone was deposited in ProOsteon without OP-1 compared to 16 (10-19) % in bone allograft without OP-1 ($p<0.05$). By adding OP-1 device to the gap, volume fraction of trabecular bone at the border of the drill-hole (zone 3) was increased from 37 (32-43)% to 44 (34-53)% ($p=0.04$). Resorption of bone allograft was significantly increased resulting in more non-mineralised tissue. No resorption of ProOsteon was detected (table III).

Table XII, study I: Bone ongrowth and gap healing.

	<i>Allograft</i>				<i>ProOsteon</i>			
	-OP-1 device		+OP-1 device		-OP-1 device		+OP-1 device	
	Zone 1	Zone 2	Zone 1	Zone 2	Zone 1	Zone 2	Zone 1	Zone 2
Bone ongrowth	13 (0-13)		13 (3-28)		12 (0-33)		39 (0-69)	
Woven bone	12 (7-20) bc	16 (10-19) bc	20 (12-27) ab	32 (24-37) ab	6 (0-13)	23 (15-26)	25 (4-35) a	26 (25-32) a
Non-mineralised tissue	54 (52-64)	60 (55-63) bc	69(68-73) abc	64 (61-75) abc	56 (51-58)	37 (34-45)	42 (39-68) a	41 (38-45)

Values reported as median (range)

a: $p<0.05$ compared to same grafting material without OP-1

b: $p<0.05$ compared to ProOsteon without OP-1

c: $p<0.05$ compared to ProOsteon with OP-1

Table XIII, study I: Fractions of grafting materials and calculation of resorption

	<i>Allograft</i>				<i>ProOsteon</i>			
	-OP-1 device		+OP-1 device		-OP-1 device		+OP-1 device	
	Zone 1	Zone 2	Zone 1	Zone 2	Zone 1	Zone 2	Zone 1	Zone 2
Time zero	33 (31-39)	36 (30-40)	29 (24-37)	28 (27-34)	39 (35-43)	40 (34-44)	31 (28-34)	32 (25-34)
3 weeks	30 (27-38)	24 (20-35) a	9 (1-18) a	2 (1-7) a	39 (36-41)	40 (39-41)	28 (24-33)	33 (28-38)
Resorption	3 (-5-7)	12 (0-16)	20 (11-28) b	28 (22-29) b	0 (-2-3)	0 (-1-1)	4 (-2-7)	0 (-7-4)

Values reported as median (range)

a: $p<0.05$ compared to time zero

b: $p<0.05$ compared to same material without OP-1

Study II

Mechanical tests

Energy absorption was significantly higher in the OP-1 treated group compared to the other three groups (table XI). The same tendency was seen in the other mechanical parameters. The implants grafted with ProOsteon alone had inferior fixation compared to the three other groups. ANOVA on ranks found no statistical difference in stiffness between the groups ($p=0.07$).

Stimulation and substitution of bone allograft around non-cemented implants

Table XIV, study II: Push-out data (median(range))

Group	Energy absorption	Ultimate shear strength	Apparent Stiffness
	(J/m ²)	(MPa)	(MPa/ mm)
Allograft, n=6	165 (59-543) ^a	1.77 (0.66-5.64)	14.0 (3.9-64.7)
ProOsteon, n=6	36 (6-306) ^b	0.44 (0.11-2.60) ^d	3.9 (0.6-18.5)
Allograft+ProOsteon, n=6	134 (16-343) ^c	1.67 (0.28-3.53)	15.3 (0.9-29.5)
Allograft+ProOsteon+OP-1, n=6	539 (202-809)	3.48 (2.01-6.19)	23.8 (3.9-58.3)

a: Allograft vs ProOsteon and allograft+ProOsteon+OP-1, p<0.05

b: ProOsteon vs allograft+ProOsteon and allograft+ProOsteon+OP-1, p<0.05

c: Allograft+ProOsteon vs allograft+ProOsteon+OP-1, p<0.05

d: ProOsteon vs allograft and allograft+ProOsteon and allograft+ProOsteon+OP-1, p<0.05

Histology

Qualitative analysis: Bone formation and bone graft resorption was mostly found in the periphery of the gap except for the OP-1 treated gaps where bone was formed and graft was resorbed also close to the implant surface (paper I, figure 3). Remnants of OP-1 collagen carrier were found in a few gaps. Resorption lacunae was recognised on the surface of allograft and ProOsteon. Non-mineralized tissue was mainly soft, cell-rich tissue with no signs of infection.

Quantitative analysis: The non-OP-1 treated implants had appr. 33% bone ingrowth whereas the OP-1 treated had ingrowth of 57% (table XV). This difference was not statistical significant (p=0.25). OP-1 device increased new bone formation significantly in zone 1 compared to the three other groups. Significantly more bone was found in zone 1 in the bone allografted group compared with the groups grafted with ProOsteon or bone allograft+ProOsteon. Only minor differences in bone formation were seen in zone 2 (table XV). The volume fraction of bone allograft after 3 weeks in the bone graft+ProOsteon group was dramatically decreased from 10 % to 2 % in zone 1 when OP-1 was added (table XVI).

Table XV, study II: Bone ingrowth and volume fractions of newly formed bone and other tissue in gap in percentage (median(range))

	Bone ingrowth	Woven bone		Soft tissue	
		Zone 1	Zone 2	Zone 1	Zone 2
		Allograft, n=6	34 (0-70)	28 (19-35) ^a	33 (30-39)
ProOsteon, n=6	33 (0-77)	20 (5-24) ^b	32 (25-39)	50 (41-72)	39 (27-42) ^b
Allograft+ProOsteon, n=6	33 (0-65)	21 (11-29) ^c	31(25-38)	51 (44-61)	38 (31-41) ^c
Allograft+ProOsteon+OP-1, n=6	57 (11-69)	32 (25-36)	33 (28-36)	56 (48-66)	46 (44-53)

a: Allograft vs ProOsteon, allograft+ProOsteon, allograft+ProOsteon+OP-1 p<0.05

b: ProOsteon vs allograft+ProOsteon+OP-1, p<0.05

c: Allograft+ProOsteon vs Allograft+ProOsteon+OP-1, p<0.05

Table XVI: Volume fractions of grafting materials in percentage (median(range)), fractions were not compared statistically

	Allograft		ProOsteon		Allograft+ProOsteon		Allograft+ProOsteon+OP-1 device	
	Zone 1	Zone 2	Zone 1	Zone 2	Zone 1	Zone 2	Zone 1	Zone 2
	Allograft (n=6)	22 (17-32)	25 (11-29)	-	-	10 (6-13)	11 (9-17)	2 (1-3)
ProOsteon (n=6)	-	-	33 (14-41)	35 (25-36)	18 (17-21)	21 (15-23)	15 (10-17)	20 (16-24)

Study III

Mechanical tests

ProOsteon alone showed inferior mechanical properties. The differences between the four groups in stiffness was not statistical significant using ANOVA on ranks ($p=0.07$). Energy absorption showed significant differences between all groups using a pairwise comparison. Eventhough ProOsteon showed high ultimate shear strength, the stiffness of ProOsteon resulted in a significant lower energy absorption compared to allograft+OP-1. However it was higher than allograft alone ($p<0.05$) (table XVII).

Table XVII, study III: Mechanical data (median (range), n=6)

Group	Ultimate shear strength (N/mm)	Energy abs. (N)	Apparent Stiffness (N/ mm ²)
Allograft	60 (6-102)	25 (1-38) ^a	190 (29-328)
ProOsteon	35 (10-57) ^a	6 (2-20) ^a	165 (11-323)
Allograft+OP-1	84 (57-109)	53 (43-81) ^a	198 (49-292)
ProOsteon+OP-1	87 (53-106)	33 (24-76) ^a	283 (61-372)

a: $p<0.05$ compared to three other groups

Study IV

Mechanical tests

Processing of bone allograft decreased all mechanical parameters however not significantly (table III). PRP had little effect on fixation (table XVIII).

Table XVIII, study IV: Push-out data (median(range))

Group	Ultimate shear strength (MPa)	Energy abs. (J/m ²)	Apparent Stiffness (MPa/mm)
Fresh frozen allograft	1.28 (0.65-2.59)	222 (91-598)	5.63 (1.9-10.7)
Processed allograft	1.20 (0.04-3.19)	213 (11-643)	5.03 (0.2-15.7)
Fresh frozen allograft+PRP	1.13 (0.17-2.63)	223 (39-382)	4.35 (0.6-7.8)
Processed allograft+PRP	1.05 (0.09-1.99)	185 (21-368)	4.29 (0.2-9.7)

None of the differences in any of the parameters were statistical significant

Histology

Qualitative analysis: There was no difference in the appearance between fresh frozen and processed bone allograft after three weeks. The HA-coating was sometimes covered with bone even when little bone formation was seen in zone 1. Tissue quantified as “non-mineralized” was mostly loose and rich in blood cells.

Stimulation and substitution of bone allograft around non-cemented implants

Quantitative analysis: The implants treated with fresh frozen bone with or without PRP had up to 59% more bone ingrowth compared to implants treated with processed bone allograft (table XIX). Also, more newly formed bone was found in zone 1 in the groups treated with fresh frozen bone allograft compared to processed bone allograft. These findings were not significant. Addition of PRP to the graft had no influence on bone ingrowth or new bone formation.

Table XIX, study IV: Bone ingrowth and volume fractions of woven bone, bone graft and non-mineralised tissue in percentages of total area (median(range))

	Bone ingrowth	Woven bone		Graft		Non-mineralised tissue	
		Zone 1	Zone 2	Zone 1	Zone 2	Zone 1	Zone 2
Fresh frozen allograft	35 (10-64)	13 (6-19)	18 (12-23)	32 (24-38)	24 (16-33)	55 (52-58)	58 (51-69)
Processed allograft	22 (8-46)	11 (6-20)	19 (9-28)	32 (27-39)	22 (14-29)	57 (50-56)	59 (50-67)
Fresh frozen allograft+PRP	30 (12-49)	15 (8-21)	21 (17-29)	28 (16-38)	22 (11-31)	58 (51-64)	57 (41-66)
Processed allograft+PRP	23 (0-58)	11 (0-19)	25 (15-33)	29 (20-40)	22 (13-32)	59 (51-69)	52 (40-63)

None of the differences in any of the parameters were statistical significant

Study V

In four of sixteen implants from the non-bone allografted groups, a preload of 2 N could not be achieved. Of those four implants, three was from the non-PRP treated (empty) group; one was from the PRP treated group.

By adding PRP to the gap, energy absorption was increased from 6 (0-14) J/m² to 14 (7-23) J/m² (NS)(table XX).

Bone allografting resulted in a 27-fold increase in ultimate strength compared to empty gap. Adding PRP to fresh frozen bone allograft did not change mechanical parameters significantly (table XX).

Table XX, study V: Push-out data (median values (interquartile ranges))

Group	Ultimate shear strength (MPa)	Energy abs. (J/m ²)	Apparent Stiffness (MPa/mm)
Empty gap	0.03 (0.00-0.04)	6 (0-14)	0.05 (0.00-0.10)
PRP	0.07 (0.03-0.13)	14 (7-38)	0.15 (0.10-0.70)
Allograft	1.59 (1.38-1.76)*	331 (209-512)*	7.50 (4.55-9.65)*
Allograft+PRP	1.56 (0.75-2.31)*	275 (88-397)*	6.95 (4.90-10.95)*

*: p<0.05 compared to no empty gap or PRP treated gaps

Histology

Qualitative analysis: In the non-bone grafted groups, woven bone was seen in the gap in both zones. The degree of bone in growth varied a lot from implant to implant, but was not associated with PRP treatment. All bone in the gap was woven. Non-mineralised tissue was mainly cell rich and bone marrow was seen. A membrane with fibres parallel to the implant surface was found in contact with implant surface in some specimen (paper V, figure 5).

In the bone grafted group, a large number of bone chips was found in both zones. They could be distinguished from newly formed bone in the lamellar appearance and also, the green colour was lighter than that of newly formed bone (figure 5). The bone chips were all cancellous bone. Newly formed bone was found in both zones and bone ongrowth into the HA coating was common.

Quantitative analysis: PRP had no influence on bone formation in the grafted or non-grafted group (table XXI). Allografting increased bone ongrowth significantly from 0 (0-8)% to 38 (33-45)% and also bone formation in both zones was increased.

Stimulation and substitution of bone allograft around non-cemented implants

Table XXI , study V: Bone ongrowth and volume fractions of woven bone, bone graft and non-mineralised tissue in percentages of total area (median values (interquartile ranges))

	Bone ongrowth	Woven bone		Graft		Non-mineralized tissue	
		Zone 1	Zone 2	Zone 1	Zone 2	Zone 1	Zone 2
Empty	0 (0-8)	13 (7-15)	15(13-18)	-	-	87(85-93)	86 (82-87)
PRP	0 (0-3)	11 (6-15)	14(11-18)	-	-	89(85-95)	87 (82-89)
Allograft	38 (33-45)*	18(14-19)*	19(15-20)*	30 (29-33)	27(23-30)	53(49-55)*	55(50-63)*
Allograft+PRP	26 (15-51)*	16(11-20)*	21(19-25)*	31 (26-34)	22 (19-25)	54(50-59)*	54(52-61)*

*: p<0.05 compared to non-allografted groups

Discussion

Fixation of implants in the ProOsteon 200 group without OP-1 was inferior to that of the bone allografted group with or without OP-1 in study I and II. This is in accordance with Turner et al¹⁵² who found limited effect of HA/TCP granules around implants compared to bone allograft. Also HA granules have shown inferior bone ingrowth in bone defect models. Mechanical evaluation of ProOsteon in the gap in study III revealed, that the inferior bone ingrowth resulted in inferior mechanical properties. Prosthesis grafted with such material might not be adequate mechanically supported. Bone formation was dramatically increased after addition of OP-1 device to ProOsteon. This is in accordance to a number of studies in which HA granules and other conductive bone substitutes have been combined with various osteoinductive agents such as BMP-3, PDGF, TGF- β or DBM at skeletal or extrasketal sites^{33,97,116}. In study III, addition of OP-1 device to ProOsteon markedly increased stiffness resulting in a statistically lower energy absorption compared to bone allograft. The impact on survival of a prosthesis grafted with HA granules with or without OP-1 in stead of bone graft is not possible to predict using our model.

ProOsteon is a slow resorbing bone substitute^{32 50,87,129} and we found no significant resorption of HA granules after three weeks not even with OP-1

Histomorphometry demonstrated an expected increase in bone formation in the bone allografted group when OP-1 was added. But furthermore we found accelerated bone graft in the OP-1 treated group. This has previously been described but not quantified¹²³. The influence of OP-1 on osteoclasts has not been investigated *in vivo* but an *in vitro* study indicates an important role of OP-1 in the recruitment of osteoclasts⁴⁸. Furthermore, preliminary results from a human trial with spinal intracorporal application of OP-1 device have shown enhanced bone resorption as a primary event⁷². Also BMP-2 stimulates osteoclasts *in vitro*⁵⁸ and PDGF has been associated with aseptical loosening of prostheses^{122,172}. In our studies on OP-1, the rate of bone graft resorption exceeded the rate of new bone formation. This mismatch lead to significant more non-mineralized tissue. Bone allograft around prostheses serves not only as a bone conductor but also provides mechanical support to the prostheses. Uncontrolled bone graft resorption prior to bone formation could theoretically lead to loss of stability of the prostheses resulting in micromotions and ultimately failure^{13,38}. OP-1 device increased bone formation in the bone allografted group but did not significantly influence implant fixation after 3 weeks. This is in accordance with previous studies where OP-1 mixed with bone allograft inserted into defects resulted in decreased mechanical fixation of HA coated or non-coated implants^{77,133}.

OP-1 device consists of OP-1 in a bovine collagen type I carrier. Our study design does not allow us to conclude, whether bone formation and graft resorption were stimulated by OP-1 or the collagen carrier. Bovine collagen type I stimulates human osteoblasts *in vitro*⁸⁹ and collagen is capable of

enhancing osteointegration of bone substitutes *in vivo*^{57,96}. However when a growth factor is added, the stimulating effect is yet markedly further improved⁹⁸.

The mechanical test of the bone grafted gaps in study III showed that ultimate shear strength and energy absorption was dramatically increased by adding OP-1 device. This indicates, that it is not the total fraction of mineralised tissue but the fraction of new bone that predicts the stability of the construct.

Tagil et al showed, that impacted graft penetrated by fibrous tissue had double compression strength compared to freshly impacted graft¹⁴⁸ and that impacted graft might not necessarily be invaded by bone to ensure mechanical stability. This study indicates, that bone ingrowth is favourable.

One question still to be answered is, if OP-1 increases bone graft resorption prior to bone formation. In that case, we might find a situation with little bone graft remained but still no new bone formation to ensure the mechanical stability. A study with more time points could answer that question.

In study IV we found no effect of processing of bone allograft. Processing by defatting, freeze drying and irradiation might have dual effects. Theoretically, bone graft processing could inactivate the growth factors and thereby delay ingrowth of bone. Growth factors contained in the graft are hypothesized to be liberated during remodelling of the graft and play an important role in bone incorporation⁸. However whereas the effect of growth factors in demineralised bone matrix is well documented, the influence of bone growth factors in mineralised bone matrix is not well elucidated. Aspenberg found only limited favourable effect of the growth factors in bone allograft when growth factors were inactivated by heat⁸. However one study has lately questioned the use of irradiated bone allograft around prosthesis since incorporation was decreased¹¹⁸.

Removal of cells and cellular debris by defatting and lavage might decrease the antigenicity of the graft and thereby increase bone ingrowth^{9,150,151}. In all studies we used a bone graft donor from the same breeder and of the same race as the recipients. We did not control if there were any antigenic mismatch between the donor of allograft and the recipient. A major mismatch might have shown differences in ingrowth between processed and fresh frozen bone allograft.

Processed bone allograft might be easier to handle and since we found no differences between fresh frozen and processed bone allograft, it should be considered in clinical practise.

In study V we found a dramatic effect of bone allografting. Søballe previously showed in a similar model, that bone grafting only had markedly effect on fixation non-HA coated but minor effect on HA coated implants after six weeks. However we showed, that the very early implant fixation can be enhanced by bone grafting.

We found no effect of PRP in study IV and V. Same dogs and PRP preparations were used in both studies. Platelets are essential in fracture repair. Platelets are activated by collagen exposure as an immediate response to fracture, leading to fibrin clotting and platelet aggregation. Platelet α -granules contain growth factors of which PDGF, TGF- β , IGF and EGF are stimulators of bone forming cells. High levels of platelets in PRP seems to be correlated with a high level of TGF- β ¹⁶⁹. The use of PRP was first described by RE Marx who increased bone incorporation of autograft in mandibular defects in humans. In rabbits PRP increases gap healing in combination with bovine cancellous bone in a calvarial defect model⁶¹ and accelerate bone incorporation of HA granules in a bone chamber¹³⁰.

We found no effect of PRP and since the activity of PRP was not tested in the present study we can not conclude if the negative result is due to inactive PRP. We reached an average platelet count of $1884 \cdot 10^9$ platelets/l in PRP which is more than the counts in our previous studies^{52,54} in canines and more than the $785 \cdot 10^9$ platelets/l in Marx study.

Marx used PRP mixed with bone graft to reconstruct large mandibular defects. In contrast, we inserted implants surrounded by a relatively small gap in young dogs with good bone healing

potential. The border of the drillhole creates a large surface from which growth factors can leak and a source of bone forming cells. We have recently shown, that platelet concentrates prepared using two different kits increases fixation of non-HA coated implants in a similar model^{52,54}. In the present study, we further optimised the gap healing potential by coating the implants with HA. HA coating is a very potent stimulator of gap healing^{138,139}. Under such optimal gap healing conditions, it might be difficult to further improve bone formation and fixation of allografted implants.

One major disadvantage of PRP is, that it might be difficult to test the quality. Each batch of bone stimulating factors such as purified BMP-2 and BMP-7 and DBM are tested by the company before it is released to the market. However that is not possible using PRP since it is processed immediately prior to operation. Since most growth factors are contained in platelets and white cell bodies it seemed logical to evaluate the quality on cell count. However only a correlation between TGF- β has been proved²⁹. Our conclusion on PRP and other platelet concentrates is hence, that experimental studies and one clinical study indicates the stimulatory effect on bone healing. However there is still a lot of work to be done to explore possible indications and to find a good way to control the level of growth factors.

Conclusion

In conclusion OP-1 device had a dual effect on incorporation of bone allograft. We found an expected increase in new bone formation however also bone graft resorption was accelerated which can explain why OP-1 did not increase fixation of bone allografted implants. The risk of failure due to accelerated resorption is already documented.

ProOsteon showed inferior bone healing capacity and can not replace bone allograft alone. However OP-1 markedly increased incorporation and fixation implants grafted with ProOsteon or a combination of ProOsteon and bone allograft.

Processing showed no impairment in bone healing. Since processing includes advantages such as easier handling and makes a safer graft to the patient, it might be considered in clinical use.

Eventhough the concept of using platelet concentrates to increase healing of bone was introduced more than 5 years ago and the concept is now commercialised, there is still little scientific evidence for the effect. We found no effect of PRP. However recent studies show, that platelet concentrates increase fixation and bone incorporation of non-HA coated titanium implants. Possible clinical applications and methods of preparation still need to be investigated.

Future research

-OP-1 markedly increased resorption of bone allograft. In a future study we will investigate, if local use of bisphosphonate block bone graft resorption.

-Local application of bisphosphonate increases incorporation of dental implants. We have planned to use local bisphosphonate to increase incorporation of titanium implants in trabecular bone.

-DBM is a source of growth factors. In a future project we will focus on a new DBM, Collos, and the influence on bone allografted implants.

-Bone marrow aspirate contain bone precursor cells. In a future project we will concentrate bone marrow aspirate and mix it with morsellized bone allograft or HA granules around titanium implants.

Reference List

1. **Aerssens, J., Boonen, S., Lowet, G., and Dequeker, J.:** Interspecies differences in bone composition, density, and quality: potential implications for in vivo bone research. *Endocrinology*. 139:663-670, 1998.
2. **Aho, A. J., Hirn, M., Aro, H. T., Heikkila, J. T., and Meurman, O.:** Bone bank service in Finland. Experience of bacteriologic, serologic and clinical results of the Turku Bone Bank 1972-1995. *Acta Orthop Scand*. 69:559-565, 1998.
3. **Anderson, M. L., Dhert, W. J., de Bruijn, J. D., Dalmeijer, R. A., Leenders, H., van Blitterswijk, C. A., and Verbout, A. J.:** Critical size defect in the goat's os ilium. A model to evaluate bone grafts and substitutes. *Clin. Orthop*. 231-239, 1999.
4. **Anitua, E.:** Plasma rich in growth factors: preliminary results of use in the preparation of future sites for implants. *Int. J. Oral Maxillofac. Implants*. 14:529-535, 1999.
5. **Arm, D. M., Tencer, A. F., Bain, S. D., and Celino, D.:** Effect of controlled release of platelet-derived growth factor from a porous hydroxyapatite implant on bone ingrowth. *Biomaterials*. 17:703-709, 1996.
6. **Aspenberg, P.:** Impaction grafting
14. *Acta Orthop. Scand*. 72:198-199, 2001.
7. **Aspenberg, P. and Astrand, J.:** Bone allografts pretreated with a bisphosphonate are not resorbed
6. *Acta Orthop. Scand*. 73:20-23, 2002.
8. **Aspenberg, P., Tagil, M., Kristensson, C., and Lidin, S.:** Bone graft proteins influence osteoconduction. A titanium chamber study in rats. *Acta Orthop. Scand*. 67:377-382, 1996.
9. **Aspenberg, P. and Thoren, K.:** Lipid extraction enhances bank bone incorporation. An experiment in rabbits. *Acta Orthop Scand*. 61:546-548, 1990.
10. **Asselmeier, M. A., Caspari, R. B., and Bottenfield, S.:** A review of allograft processing and sterilization techniques and their role in transmission of the human immunodeficiency virus. *Am. J. Sports Med*. 21:170-175, 1993.
11. **Berzins, A., Sumner, D. R., Wasielewski, R. C., and Galante, J. O.:** Impacted particulate allograft for femoral revision total hip arthroplasty. In vitro mechanical stability and effects of cement pressurization. *J Arthroplasty*. 11:500-506, 1996.
12. **Blom, A. W., Grimm, B., Miles, A. W., Cunningham, J. L., and Learmonth, I. D.:** Subsidence in impaction grafting: the effect of adding a ceramic bone graft extender to bone. *Proc. Inst. Mech. Eng [H.]*. 216:265-270, 2002.
13. **Bostrom, M., Lane, J. M., Tomin, E., Browne, M., Berberian, W., Turek, T., Smith, J., Wozney, J., and Schildhauer, T.:** Use of bone morphogenetic protein-2 in the rabbit ulnar nonunion model. *Clin. Orthop*. 272-282, 1996.
14. **Brodt, M. D., Swan, C. C., and Brown, T. D.:** Mechanical behavior of human morselized cancellous bone in triaxial compression testing. *J. Orthop. Res*. 16:43-49, 1998.

Stimulation and substitution of bone allograft around non-cemented implants

15. **Bucholz, R. W., Carlton, A., and Holmes, R. E.:** Hydroxyapatite and tricalcium phosphate bone graft substitutes. *Orthop. Clin. North Am.* 18:323-334, 1987.
16. **Bujia, J., Wilmes, E., Kastenbauer, E., and Gurtler, L.:** Influence of chemical allograft preservation procedures on the human immunodeficiency virus. *Laryngoscope.* 106:645-647, 1996.
17. **Buma, P., Lamerigts, N., Schreurs, B. W., Gardeniers, J., Versleyen, D., and Slooff, T. J.:** Impacted graft incorporation after cemented acetabular revision. Histological evaluation in 8 patients. *Acta Orthop Scand.* 67:536-540, 1996.
18. **Cacciafesta, V., Dalstra, M., Bosch, C., Melsen, B., and Andreassen, T. T.:** Growth hormone treatment promotes guided bone regeneration in rat calvarial defects 3. *Eur. J. Orthod.* 23:733-740, 2001.
19. **Conrad, E. U., Ericksen, D. P., Tencer, A. F., Strong, D. M., and Mackenzie, A. P.:** The effects of freeze-drying and rehydration on cancellous bone. *Clin Orthop.* 290:279-84, 1993.
20. **Cook, S. D., Baffes, G. C., Wolfe, M. W., Sampath, T. K., and Rueger, D. C.:** Recombinant human bone morphogenetic protein-7 induces healing in a canine long-bone segmental defect model. *Clin. Orthop.* 302-312, 1994.
21. **Cook, S. D., Baffes, G. C., Wolfe, M. W., Sampath, T. K., Rueger, D. C., and Whitecloud, T. S.:** The effect of recombinant human osteogenic protein-1 on healing of large segmental bone defects. *J. Bone Joint Surg. Am.* 76:827-838, 1994.
22. **Cook, S. D., Salkeld, S. L., Patron, L. P., and Barrack, R. L.:** The effect of demineralized bone matrix gel on bone ingrowth and fixation of porous implants 1. *J. Arthroplasty.* 17:402-408, 2002.
23. **Cook, S. D., Salkeld, S. L., and Rueger, D. C.:** Evaluation of recombinant human osteogenic protein-1 (rhOP-1) placed with dental implants in fresh extraction sites. *J. Oral Implantol.* 21:281-289, 1995.
24. **Cornu, O., Banse, X., Docquier, P. L., Luyckx, S., and Delloye, C.:** Effect of freeze-drying and gamma irradiation on the mechanical properties of human cancellous bone. *J Orthop Res.* 18:426-31, 2000.
25. **Costantino, P. D. and Friedman, C. D.:** Synthetic bone graft substitutes. *Otolaryngol. Clin. North Am.* 27:1037-1074, 1994.
26. **De Roeck, N. J. and Drabu, K. J.:** Impaction bone grafting using freeze-dried allograft in revision hip arthroplasty. *J Arthroplasty.* 16:201-6, 2001.
27. **den Boer, F. C., Wippermann, B. W., Blokhuis, T. J., Patka, P., Bakker, F. C., and Haarman, H. J.:** Healing of segmental bone defects with granular porous hydroxyapatite augmented with recombinant human osteogenic protein-1 or autologous bone marrow. *J. Orthop. Res.* 21:521-528, 2003.
28. **Dhert, W. J., Thomsen, P., Blomgren, A. K., Esposito, M., Ericson, L. E., and Verbout, A. J.:** Integration of press-fit implants in cortical bone: a study on interface kinetics. *J. Biomed. Mater. Res.* 41:574-583, 1998.
29. **Dugrillon, A., Eichler, H., Kern, S., and Kluter, H.:** Autologous concentrated platelet-rich plasma (cPRP) for local application in bone regeneration. *Int. J. Oral Maxillofac. Surg.* 31:615-619, 2002.
30. **Eby, B. W.:** Platelet-rich plasma: harvesting with a single-spin centrifuge. *J. Oral Implantol.* 28:297-301, 2002.
31. **Eggl, P. S., Muller, W., and Schenk, R. K.:** Porous hydroxyapatite and tricalcium phosphate cylinders with two different pore size ranges implanted in the cancellous bone of rabbits. A comparative histomorphometric and histologic study of bony ingrowth and implant substitution. *Clin. Orthop.* 127-138, 1988.

Stimulation and substitution of bone allograft around non-cemented implants

32. **el Deeb, M. and Holmes, R. E.:** Zygomatic and mandibular augmentation with proplast and porous hydroxyapatite in rhesus monkeys. *J. Oral Maxillofac. Surg.* 47:480-488, 1989.
33. **el Deeb, M., Hosny, M., and Sharawy, M.:** Osteogenesis in composite grafts of allogenic demineralized bone powder and porous hydroxylapatite. *J. Oral Maxillofac. Surg.* 47:50-56, 1989.
34. **Eldridge, J. D., Cunningham, J. L., Samuels, A., Blunn, G. W., Lawes, T. J., Learmonth, I. D., and Goodship, A. E.:** Glass ionomer as an expander of allograft in revision arthroplasty of the hip. *Biomaterials.* 24:499-508, 2003.
35. **Eldridge, J. D., Smith, E. J., Hubble, M. J., Whitehouse, S. L., and Learmonth, I. D.:** Massive early subsidence following femoral impaction grafting. *J Arthroplasty.* 12:535-540, 1997.
36. **Elting, J. J., Mikhail, W. E., Zicat, B. A., Hubbell, J. C., Lane, L. E., and House, B.:** Preliminary report of impaction grafting for exchange femoral arthroplasty. *Clin Orthop.*159-167, 1995.
37. **Enneking, W. F. and Mindell, E. R.:** Observations on massive retrieved human allografts. *J Bone Joint Surg Am.* 73:1123-1142, 1991.
38. **Fischgrund, J. S., James, S. B., Chabot, M. C., Hankin, R., Herkowitz, H. N., Wozney, J. M., and Shirkhoda, A. :** Augmentation of autograft using rhBMP-2 and different carrier media in the canine spinal fusion model. *J. Spinal. Disord.* 10:467-472, 1997.
39. **Friedlaender, G. E., Perry, C. R., Cole, J. D., Cook, S. D., Cierny, G., Muschler, G. F., Zych, G. A., Calhoun, J. H., LaForte, A. J., and Yin, S.:** Osteogenic protein-1 (bone morphogenetic protein-7) in the treatment of tibial nonunions. *J. Bone Joint Surg. Am. suppl* 1:S151-8, 2001.
40. **Galea, G., Kopman, D., and Graham, B. J.:** Supply and demand of bone allograft for revision hip surgery in Scotland. *J. Bone Joint Surg. Br.* 80:595-599, 1998.
41. **Gie, G. A., Linder, L., Ling, R. S., Simon, J. P., Slooff, T. J., and Timperley, A. J.:** Impacted cancellous allografts and cement for revision total hip arthroplasty. *J. Bone Joint Surg. Br.* 75:14-21, 1993.
42. **Giesen, E. B., Lamerigts, N. M., Verdonshot, N., Buma, P., Schreurs, B. W., and Huiskes, R.:** Mechanical characteristics of impacted morsellised bone grafts used in revision of total hip arthroplasty. *J Bone Joint Surg Br.* 81:1052-1057, 1999.
43. **Goldberg, V. M., Bos, G. D., Heiple, K. G., Zika, J. M., and Powell, A. E.:** Improved acceptance of frozen bone allografts in genetically mismatched dogs by immunosuppression. *J. Bone Joint Surg. Am.* 66:937-950, 1984.
44. **Goldberg, V. M., Powell, A., Shaffer, J. W., Zika, J., Bos, G. D., and Heiple, K. G.:** Bone grafting: role of histocompatibility in transplantation. *J. Orthop. Res.* 3:389-404, 1985.
45. **Gotfredsen, K., Budtz-Jørgensen, E., and Jensen, L. L.:** Preparation and staining of sections containing titanium-implants. *Stain Tecnology.* 64:121-127, 1989.
46. **Hamadouche, M., Blanchat, C., Meunier, A., Kerboull, L., and Kerboull, M.:** Histological findings in a proximal femoral structural allograft ten years following revision total hip arthroplasty: a case report. *J. Bone Joint Surg. Am.* 84-A:269-273, 2002.
47. **Henman, P. and Finlayson, D.:** Ordering allograft by weight: suggestions for the efficient use of frozen bone-graft for impaction grafting. *J Arthroplasty.* 15:368-371, 2000.
48. **Hentunen, T. A., Lakkakorpi, P. T., Tuukkanen, J., Lehenkari, P. P., Sampath, T. K., and Vaananen, H. K.:** Effects of recombinant human osteogenic protein-1 on the differentiation of osteoclast-like cells and bone resorption. *Biochem. Biophys. Res. Commun.* 209:433-443, 1995.

Stimulation and substitution of bone allograft around non-cemented implants

49. **Herberts, P. and Malchau, H.:** How outcome studies have changed total hip arthroplasty practices in Sweden. *Clin. Orthop.*44-60, 1997.
50. **Holmes, R. E., Bucholz, R. W., and Mooney, V.:** Porous hydroxyapatite as a bone graft substitute in diaphyseal defects: a histometric study. *J. Orthop. Res.* 5:114-121, 1987.
51. **Huja, S. S., Katona, T. R., Burr, D. B., Garetto, L. P., and Roberts, W. E.:** Microdamage adjacent to endosseous implants
2. *Bone.* 25:217-222, 1999.
52. **Jensen, T. B., Bechtold, J. E., Chen, X., Elmengaard, B., and Soballe, K.** Platelet concentrate improves fixation achieved with morselized bone allograft around non-cemented implants. *Trans.49th Annual Meeting Orthop Res .* 2003.
Ref Type: Abstract
53. **Jensen, T. B., Overgaard, S., Lind, M., Rahbek, O., Bunger, C., and Soballe, K.:** Osteogenic protein 1 device increases bone formation and bone graft resorption around cementless implants. *Acta Orthop. Scand.* 73:31-39, 2002.
54. **Jensen, T., Bechtold, J. E., Cheng, X., Kidder, L., and Soballe, K.:** Autologous Growth Factor (AGF®) in combination with morselized bone allograft improves implant fixation. *Trans. 48th Annual Meeting, ORS.* 2002.
55. **Jeppsson, C, Wang, J, Tagil, M., and Aspenberg, P.** No augmentation by OP-1 in compacted bone allograft around rabbit knee prostheses. *Trans.47th Annual Meeting, ORS ,* 0592. 2001.
Ref Type: Abstract
56. **Jinno, T., Miric, A., Feighan, J., Kirk, S. K., Davy, D. T., and Stevenson, S.:** The effects of processing and low dose irradiation on cortical bone grafts. *Clin Orthop.*275-85, 2000.
57. **Johnson, K. D., Frierson, K. E., Keller, T. S., Cook, C., Scheinberg, R., Zerwekh, J., Meyers, L., and Sciadini, M. F.:** Porous ceramics as bone graft substitutes in long bone defects: a biomechanical, histological, and radiographic analysis. *J. Orthop. Res.* 14:351-369, 1996.
58. **Kanatani, M., Sugimoto, T., Kaji, H., Kobayashi, T., Nishiyama, K., Fukase, M., Kumegawa, M., and Chihara, K.:** Stimulatory effect of bone morphogenetic protein-2 on osteoclast-like cell formation and bone-resorbing activity. *J. Bone Miner. Res.* 10:1681-1690, 1995.
59. **Karrholm, J., Borssen, B., Lowenhielm, G., and Snorrason, F.:** Does early micromotion of femoral stem prostheses matter? 4-7-year stereoradiographic follow-up of 84 cemented prostheses. *J Bone Joint Surg Br.* 76:912-7, 1994.
60. **Karrholm, J. and Snorrason, F. :** Subsidence, tip, and hump micromovements of noncoated ribbed femoral prostheses. *Clin Orthop.*50-60, 1993.
61. **Kim, E. S., Park, E. J., and Choung, P. H.:** Platelet concentration and its effect on bone formation in calvarial defects: An experimental study in rabbits. *J Prosthet. Dent.* 86:428-433, 2001.
62. **Kim, S. G., Chung, C. H., Kim, Y. K., Park, J. C., and Lim, S. C.:** Use of particulate dentin-plaster of Paris combination with/without platelet-rich plasma in the treatment of bone defects around implants
1. *Int. J. Oral Maxillofac. Implants.* 17:86-94, 2002.
63. **Kim, S. G., Kim, W. K., Park, J. C., and Kim, H. J.:** A comparative study of osseointegration of Avana implants in a demineralized freeze-dried bone alone or with platelet-rich plasma. *J. Oral Maxillofac. Surg.* 60:1018-1025, 2002.
64. **Kimmel, D. B. and Jee, W. S.:** A quantitative histologic study of bone turnover in young adult beagles. *Anat. Rec.* 203:31-45, 1982.

Stimulation and substitution of bone allograft around non-cemented implants

65. **Klinger, M. H. and Jelkmann, W.:** Role of blood platelets in infection and inflammation. *J. Interferon Cytokine Res.* 22:913-922, 2002.
66. **Knighton, D. R., Ciresi, K. F., Fiegel, V. D., Austin, L. L., and Butler, E. L.:** Classification and treatment of chronic nonhealing wounds. Successful treatment with autologous platelet-derived wound healing factors (PDWHF). *Ann. Surg.* 204:322-330, 1986.
67. **Krupski, W. C., Reilly, L. M., Perez, S., Moss, K. M., Crombleholme, P. A., and Rapp, J. H.:** A prospective randomized trial of autologous platelet-derived wound healing factors for treatment of chronic nonhealing wounds: a preliminary report. *J Vasc. Surg.* 14:526-532, 1991.
68. **Lamerigts, N., Aspenberg, P., Buma, P., Versleyen, D., and Slooff, T. J.:** The repeated sampling bone chamber: a new permanent titanium implant to study bone grafts in the goat. *Lab. Anim. Sci.* 47:401-406, 1997.
69. **Lamerigts, N. M., Buma, P., Aspenberg, P., Schreurs, B. W., and Slooff, T. J.:** Role of growth factors in the incorporation of unloaded bone allografts in the goat. *Clin Orthop.* 260-270, 1999.
70. **Lamerigts, N. M., Buma, P., Huiskes, R., Schreurs, W., Gardeniers, J., and Slooff, T. J.:** Incorporation of morsellized bone graft under controlled loading conditions. A new animal model in the goat. *Biomaterials.* 21:741-747, 2000.
71. **Landesberg, R., Roy, M., and Glickman, R. S.:** Quantification of growth factor levels using a simplified method of platelet-rich plasma gel preparation. *J. Oral Maxillofac. Surg.* 58:297-300, 2000.
72. **Laursen, M., Hoey, K., Hansen, E. S., Gelineck, J., Christensen, F., and Bunger, C.:** Recombinant bone morphogenic protein-7 as an intracorporal bone growth stimulator in unstable thoracolumbar burst fractures in humans; preliminary results. *European Spine Journal.* 8:485-490, 1999.
73. **Lind, M., Overgaard, S., Jensen, T. B., Song, Y., Goodman, S. B., Bunger, C., and Soballe, K.:** Effect of osteogenic protein 1/collagen composite combined with impacted allograft around hydroxyapatite-coated titanium alloy implants is moderate. *J Biomed. Mater. Res.* 55:89-95, 2001.
74. **Lind, M., Overgaard, S., Nguyen, T., Ongpipattanakul, B., Bunger, C., and Soballe, K.:** Transforming growth factor-beta stimulates bone ongrowth. Hydroxyapatite-coated implants studied in dogs. *Acta Orthop. Scand.* 67:611-616, 1996.
75. **Lind, M., Overgaard, S., Ongpipattanakul, B., Nguyen, T., Bunger, C., and Soballe, K.:** Transforming growth factor-beta 1 stimulates bone ongrowth to weight-loaded tricalcium phosphate coated implants: an experimental study in dogs. *J. Bone Joint Surg. Br.* 78:377-382, 1996.
76. **Lind, M., Overgaard, S., Soballe, K., Nguyen, T., Ongpipattanakul, B., and Bunger, C.:** Transforming growth factor-beta 1 enhances bone healing to unloaded tricalcium phosphate coated implants: an experimental study in dogs. *J. Orthop. Res.* 14:343-350, 1996.
77. **Lind, M., Overgaard, S., Song, Y., Jensen, T., Goodman, S., Büngrer, C., and Soballe, K.:** Osteogenic Protein 1 enhances mechanical fixation of implants in trabecular bone. *Trans. 44th Annual Meeting ORS.* 339, 1998.
78. **Lind, M., Overgaard, S., Song, Y., Goodman, S. B., Bunger, C., and Soballe, K.:** Osteogenic protein 1 device stimulates bone healing to hydroxyapatite-coated and titanium implants. *J Arthroplasty.* 15:339-46, 2000.
79. **Linder, L.:** Cancellous impaction grafting in the human femur: histological and radiographic observations in 6 autopsy femurs and 8 biopsies. *Acta Orthop Scand.* 71:543-552, 2000.
80. **Ling, R. S.:** Femoral reconstruction with morcelized bone graft and cemented stem. *Chir Organi Mov.* 79:305-311, 1994.
81. **Lucht, U.:** The Danish Hip Arthroplasty Register. *Acta Orthop. Scand.* 71:433-439, 2000.

Stimulation and substitution of bone allograft around non-cemented implants

82. **Lucht, U., Bungler, C., Moller, J. T., Joyce, F., and Plenk, H.:** Fibrin sealant in bone transplantation. No effects on blood flow and bone formation in dogs. *Acta Orthop Scand.* 57:19-24, 1986.
83. **Malkani, A. L., Voor, M. J., Fee, K. A., and Bates, C. S.:** Femoral component revision using impacted morsellised cancellous graft. A biomechanical study of implant stability [see comments]. *J. Bone Joint Surg. Br.* 78:973-978, 1996.
84. **Man, D., Plosker, H., and Winland-Brown, J. E.:** The use of autologous platelet-rich plasma (platelet gel) and autologous platelet-poor plasma (fibrin glue) in cosmetic surgery. *Plast. Reconstr. Surg.* 107:229-237, 2001.
85. **Mann, C. J., McNally, S., Taylor, E., and Shepperd, J. A.:** A retrospective clinical and radiographic review of 173 hydroxyapatite-coated screw cups with 5- to 10-year follow-up, showing low revision rates for fixation failure. *J. Arthroplasty.* 17:851-855, 2002.
86. **Marthy, S. and Richter, M.:** Human immunodeficiency virus activity in rib allografts. *J. Oral Maxillofac. Surg.* 56:474-476, 1998.
87. **Martin, R. B., Chapman, M. W., Sharkey, N. A., Zissimos, S. L., Bay, B., and Shors, E. C.:** Bone ingrowth and mechanical properties of coralline hydroxyapatite 1 yr after implantation. *Biomaterials.* 14:341-348, 1993.
88. **Marx, R. E., Carlson, E. R., Eichstaedt-RM, Schimmele-SR, Strauss-JE, and Georgeff-KR:** Platelet-rich plasma: Growth factor enhancement for bone grafts. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endod.* 85:638-646, 1998.
89. **Masi, L., Franchi, A., Santucci, M., Danielli, D., Arganini, L., Giannone, V., Formigli, L., Benvenuti, S., Tanini, A., Beghe, F., and et al:** Adhesion, growth, and matrix production by osteoblasts on collagen substrata. *Calcif. Tissue Int.* 51:202-212, 1992.
90. **McDonald, D. J., Fitzgerald, R. H., Jr., and Chao, E. Y.:** The enhancement of fixation of a porous-coated femoral component by autograft and allograft in the dog. *J. Bone Joint Surg. Am.* 70:728-737, 1988.
91. **Meding, J. B., Ritter, M. A., Keating, E. M., and Faris, P. M.:** Impaction bone-grafting before insertion of a femoral stem with cement in revision total hip arthroplasty. A minimum two-year follow-up study. *J Bone Joint Surg Am.* 79:1834-1841, 1997.
92. **Mulroy, W. F. and Harris, W. H.:** Revision total hip arthroplasty with use of so-called second-generation cementing techniques for aseptic loosening of the femoral component. A fifteen-year-average follow-up study. *J. Bone Joint Surg. Am.* 78:325-330, 1996.
93. **Nelissen, R. G., Bauer, T. W., Weidenhielm, L. R., LeGolvan, D. P., and Mikhail, W. E.:** Revision hip arthroplasty with the use of cement and impaction grafting. Histological analysis of four cases. *J. Bone Joint Surg. Am.* 77:412-422, 1995.
94. **Nelissen, R. G., Valstar, E. R., Poll, R. G., Garling, E. H., and Brand, R.:** Factors associated with excessive migration in bone impaction hip revision surgery: A radiostereometric analysis study. *J. Arthroplasty.* 17:826-833, 2002.
95. **Nemzek, J. A., Arnoczky, S. P., and Swenson, C. L.:** Retroviral transmission by the transplantation of connective-tissue allografts. An experimental study. *J. Bone Joint Surg. Am.* 76:1036-1041, 1994.
96. **Ono, I., Gunji, H., Kaneko, F., Saito, T., and Kuboki, Y.:** Efficacy of hydroxyapatite ceramic as a carrier for recombinant human bone morphogenetic protein. *J. Craniofac. Surg.* 6:238-244, 1995.
97. **Ono, I., Inoue, M., and Kuboki, Y.:** Promotion of the osteogenetic activity of recombinant human bone morphogenetic protein by prostaglandin E1. *Bone.* 19:581-588, 1996.

Stimulation and substitution of bone allograft around non-cemented implants

98. **Ono, I., Ohura, T., Murata, M., Yamaguchi, H., Ohnuma, Y., and Kuboki, Y.:** A study on bone induction in hydroxyapatite combined with bone morphogenetic protein. *Plast. Reconstr. Surg.* 90:870-879, 1992.
99. **Oonishi, H., Hench, L. L., Wilson, J., Sugihara, F., Tsuji, E., Matsuura, M., Kin, S., Yamamoto, T., and Mizokawa, S.:** Quantitative comparison of bone growth behavior in granules of Bioglass, A-W glass-ceramic, and hydroxyapatite
J. Biomed. Mater. Res. 51:37-46, 2000.
100. **Oonishi, H., Iwaki, Y., Kin, N., Kushitani, S., Murata, N., Wakitani, S., and Imoto, K.:** Hydroxyapatite in revision of total hip replacements with massive acetabular defects: 4- to 10-year clinical results. *J. Bone Joint Surg. Br.* 79:87-92, 1997.
101. **Oonishi, H., Kushitani, S., Yasukawa, E., Iwaki, H., Hench, L. L., Wilson, J., Tsuji, E., and Sugihara, T.:** Particulate bioglass compared with hydroxyapatite as a bone graft substitute
Clin. Orthop. 316-325, 1997.
102. **Overgaard, S.:** Calcium phosphate coatings for fixation of bone implants-Evaluated mechanically and histologically by stereological methods. *Acta Orthop. Scand.* 71: 2000.
103. **Overgaard, S., Lind, M., Glerup, H., Grundvig, S., Bunger, C., and Soballe, K.:** Hydroxyapatite and fluorapatite coatings for fixation of weight loaded implants. *Clin. Orthop.* 286-296, 1997.
104. **Overgaard, S., Lind, M., Josephsen, K., Maunsbach, A. B., Bunger, C., and Soballe, K.:** Resorption of hydroxyapatite and fluorapatite ceramic coatings on weight-bearing implants: a quantitative and morphological study in dogs. *J. Biomed. Mater. Res.* 39:141-152, 1998.
105. **Overgaard, S., Lind, M., Rahbek, O., Bunger, C., and Soballe, K.:** Improved fixation of porous-coated versus grit-blasted surface texture of hydroxyapatite-coated implants in dogs. *Acta Orthop. Scand.* 68:337-343, 1997.
106. **Overgaard, S., Soballe, K., Jorgen, H., and Gundersen, G.:** Efficiency of systematic sampling in histomorphometric bone research illustrated by hydroxyapatite-coated implants: optimizing the stereological vertical-section design. *J Orthop Res.* 18:313-321, 2000.
107. **Overgaard, S., Soballe, K., Josephsen, K., Hansen, E. S., and Bunger, C.:** Role of different loading conditions on resorption of hydroxyapatite coating evaluated by histomorphometric and stereological methods. *J. Orthop. Res.* 14:888-894, 1996.
108. **Pallesen, L., Schou, S., Aaboe, M., Hjorting-Hansen, E., Nattestad, A., and Melsen, F.:** Influence of particle size of autogenous bone grafts on the early stages of bone regeneration: a histologic and stereologic study in rabbit calvarium. *Int. J. Oral Maxillofac. Implants.* 17:498-506, 2002.
109. **Palmer, S. H., Gibbons, C. L., and Athanasou, N. A.:** The pathology of bone allograft. *J. Bone Joint Surg. Br.* 81:333-335, 1999.
110. **Pekkarinen, J., Alho, A., Lepisto, J., Ylikoski, M., Ylinen, P., and Paavilainen, T.:** Impaction bone grafting in revision hip surgery. A high incidence of complications. *J. Bone Joint Surg. Br.* 82:103-107, 2000.
111. **Pratt, J. N., Griffon, D. J., Dunlop, D. G., Smith, N., and Howie, C. R.:** Impaction grafting with morsellised allograft and tricalcium phosphate- hydroxyapatite: incorporation within ovine metaphyseal bone defects. *Biomaterials.* 23:3309-3317, 2002.
112. **Rahbek, O., Overgaard, S., Jensen, T. B., Bendix, K., and Soballe, K.:** Sealing effect of hydroxyapatite coating: a 12-month study in canines. *Acta Orthop Scand.* 71:563-73, 2000.
113. **Rahbek, O., Overgaard, S., Lind, M., Bendix, K., Bunger, C., and Soballe, K.:** Sealing effect of hydroxyapatite coating on peri-implant migration of particles. An experimental study in dogs. *J Bone Joint Surg Br.* 83:441-447, 2001.

Stimulation and substitution of bone allograft around non-cemented implants

114. **Randall, R. L., Pelker, R. R., Friedlaender, G. E., Goldsmith, S. L., and Panjabi, M. M.:** Sequential dependence of freeze-drying and irradiation on biomechanical properties of rat bone. *Am. J. Orthop.* 31:129-134, 2002.
115. **Retpen, J. B., Varmarken, J. E., Rock, N. D., and Jensen, J. S.:** Unsatisfactory results after repeated revision of hip arthroplasty. 61 cases followed for 5 (1-10) years. *Acta Orthop. Scand.* 63:120-127, 1992.
116. **Ripamonti, U., Ma, S. S., and Reddi, A. H.:** Induction of bone in composites of osteogenin and porous hydroxyapatite in baboons. *Plast. Reconstr. Surg.* 89:731-739, 1992.
117. **Ripamonti, U., Ma, S. S., Van den Heever, B., and Reddi, A. H.:** Osteogenin, a bone morphogenetic protein, adsorbed on porous hydroxyapatite substrata, induces rapid bone differentiation in calvarial defects of adult primates. *Plast. Reconstr. Surg.* 90:382-393, 1992.
118. **Robinson, D. E., Lee, M. B., Smith, E. J., and Learmonth, I. D.:** Femoral impaction grafting in revision hip arthroplasty with irradiated bone. *J. Arthroplasty.* 17:834-840, 2002.
119. **Rokkum, M., Brandt, M., Bye, K., Hetland, K. R., Waage, S., and Reigstad, A.:** Polyethylene wear, osteolysis and acetabular loosening with an HA- coated hip prosthesis. A follow-up of 94 consecutive arthroplasties. *J. Bone Joint Surg. Br.* 81:582-589, 1999.
120. **Roux, F. X., Brasnu, D., Menard, M., Devaux, B., Nohra, G., and Loty, B.:** Madreporic coral for cranial base reconstruction. 8years experience. . *Acta Neurochir. Wien.* 133:201-205, 1995.
121. **Rutherford, R. B., Sampath, T. K., Rueger, D. C., and Taylor, T. D.:** Use of bovine osteogenic protein to promote rapid osseointegration of endosseous dental implants. *Int. J. Oral Maxillofac. Implants.* 7:297-301, 1992.
122. **Salcetti, J. M., Moriarty, J. D., Cooper, L. F., Smith, F. W., Collins, J. G., Socransky, S. S., and Offenbacher, S.:** The clinical, microbial, and host response characteristics of the failing implant. *Int. J. Oral Maxillofac. Implants.* 12:32-42, 1997.
123. **Salkeld, S. L., Rueger, D. C., Popich L.S., and Cook, S. D.:** Improved performance of autograft and allograft bone with Osteogenic Protein 1. *Trans. 43rd Annual Meeting ORS.*255, 1997.
124. **Salzman, N. P., Psallidopoulos, M., Prewett, A. B., and O'Leary, R.:** Detection of HIV in bone allografts prepared from AIDS autopsy tissue. *Clin. Orthop.*384-390, 1993.
125. **Sampath, T. K., Coughlin, J. E., Whetstone, R. M., Banach, D., Corbett, C., Ridge, R. J., Ozkaynak, E., Oppermann, H., and Rueger, D. C.:** Bovine osteogenic protein is composed of dimers of OP-1 and BMP- 2A, two members of the transforming growth factor-beta superfamily. *J. Biol. Chem.* 265:13198-13205, 1990.
126. **Schreurs, B. W., Buma, P., Huiskes, R., Slagter, J. L., and Slooff, T. J.:** Morsellized allografts for fixation of the hip prosthesis femoral component. A mechanical and histological study in the goat. *Acta Orthop. Scand.* 65:267-275, 1994.
127. **Schreurs, B. W., Huiskes, R., Buma, P., and Slooff, T. J.:** Biomechanical and histological evaluation of a hydroxyapatite-coated titanium femoral stem fixed with an intramedullary morsellized bone grafting technique: an animal experiment on goats. *Biomaterials.* 17:1177-1186, 1996.
128. **Shanbhag, A. S., Hasselman, C. T., and Rubash, H. E.:** The John Charnley Award. Inhibition of wear debris mediated osteolysis in a canine total hip arthroplasty model. *Clin Orthop.*33-43, 1997.
129. **Shimazaki, K. and Mooney, V.:** Comparative study of porous hydroxyapatite and tricalcium phosphate as bone substitute. *J. Orthop. Res.* 3:301-310, 1985.

Stimulation and substitution of bone allograft around non-cemented implants

130. **Siebrecht, M. A., De Rooij, P. P., Arm, D. M., Olsson, M. L., and Aspenberg, P.:** Platelet concentrate increases bone ingrowth into porous hydroxyapatite
9. Orthopedics. 25:169-172, 2002.
131. **Slater, M., Patava, J., Kingham, K., and Mason, R. S.:** Involvement of platelets in stimulating osteogenic activity. J Orthop Res. 13:655-663, 1995.
132. **Slooff, T. J., Huiskes, R., van Horn, J., and Lemmens, A. J.:** Bone grafting in total hip replacement for acetabular protrusion. Acta Orthop. Scand. 55:593-596, 1984.
133. **Soballe, K. and Bechtold, J.:** Differential response to OP-1 in primary and revision implants. Trans. 45th Annual Meeting, ORS. 1999.
134. **Soballe, K.:** Hydroxyapatite ceramic coating for bone implant fixation. Mechanical and histological studies in dogs. Acta Orthop. Scand. Suppl. 255, vol.64:1-58, 1993.
135. **Soballe, K., Brockstedt Rasmussen, H., Hansen, E. S., and Bunger, C.:** Hydroxyapatite coating modifies implant membrane formation. Controlled micromotion studied in dogs. Acta Orthop. Scand. 63:128-140, 1992.
136. **Soballe, K., Hansen, E. S., B.Rasmussen, H., Jorgensen, P. H., and Bunger, C.:** Tissue ingrowth into titanium and hydroxyapatite-coated implants during stable and unstable mechanical conditions. J. Orthop. Res. 10:285-299, 1992.
137. **Soballe, K., Hansen, E. S., Brockstedt-Rasmussen, H., Hjortdal, V. E., Juhl, G. I., Pedersen, C. M., Hvid, I., and Bunger, C.:** Fixation of titanium and hydroxyapatite-coated implants in arthritic osteopenic bone. J. Arthroplasty. 6:307-316, 1991.
138. **Soballe, K., Hansen, E. S., Brockstedt Rasmussen, H., and Bunger, C.:** Hydroxyapatite coating converts fibrous tissue to bone around loaded implants. J. Bone Joint Surg. Br. 75:270-278, 1993.
139. **Soballe, K., Hansen, E. S., Brockstedt Rasmussen, H., Pedersen, C. M., and Bunger, C.:** Bone graft incorporation around titanium-alloy- and hydroxyapatite-coated implants in dogs. Clin. Orthop.282-293, 1992.
140. **Soballe, K. and Overgaard, S. :** The current status of hydroxyapatite coating of prostheses [editorial]. J. Bone Joint Surg. Br. 78:689-691, 1996.
141. **Sommerville, S. M., Johnson, N., Bryce, S. L., Journeaux, S. F., and Morgan, D. A.:** Contamination of banked femoral head allograft: incidence, bacteriology and donor follow up. Aust. N. Z. J. Surg. 70:480-484, 2000.
142. **Steed, D. L.:** The role of growth factors in wound healing. Surg. Clin. North Am. 77:575-586, 1997.
143. **Steinberg, L. A.:** The omnipotent platelet. Med. Hypotheses. 46:188-190, 1996.
144. **Stevenson, S., Li, X. Q., Davy, D. T., Klein, L., and Goldberg, V. M.:** Critical biological determinants of incorporation of non-vascularized cortical bone grafts. Quantification of a complex process and structure. J Bone Joint Surg Am. 79:1-16, 1997.
145. **Sumner, D. R., Turner, T. M., Purchio, A. F., Gombotz, W. R., Urban, R. M., and Galante, J. O.:** Enhancement of bone ingrowth by transforming growth factor-beta. J. Bone Joint Surg. Am. 77:1135-1147, 1995.
146. **Sutherland, A. G., Raafat, A., Yates, P., and Hutchison, J. D.:** Infection associated with the use of allograft bone from the north east Scotland Bone Bank. J. Hosp. Infect. 35:215-222, 1997.
147. **Tagil, M. and Aspenberg, P.:** Impaction of cancellous bone grafts impairs osteoconduction in titanium chambers. Clin Orthop.231-238, 1998.

Stimulation and substitution of bone allograft around non-cemented implants

148. **Tagil, M. and Aspenberg, P.:** Fibrous tissue armoring increases the mechanical strength of an impacted bone graft. *Acta Orthop Scand.* 72:78-82, 2001.
149. **Tayapongsak, P., O'Brien, D. A., Monteiro, C. B., and Arceo, D. L.:** Autologous fibrin adhesive in mandibular reconstruction with particulate cancellous bone and marrow. *J. Oral Maxillofac. Surg.* 52:161-165, 1994.
150. **Thoren, K., Aspenberg, P., and Thorngren, K. G.:** Lipid extraction decreases the specific immunologic response to bone allografts in rabbits. *Acta Orthop Scand.* 64:44-6, 1993.
151. **Thoren, K., Aspenberg, P., and Thorngren, K. G.:** Lipid extracted bank bone. Bone conductive and mechanical properties. *Clin Orthop.* 232-46, 1995.
152. **Turner, T. M., Urban, R. M., Sumner, D. R., and Galante, J. O.:** Revision, without cement, of aseptically loose, cemented total hip prostheses. Quantitative comparison of the effects of four types of medullary treatment on bone ingrowth in a canine model [see comments]. *J. Bone Joint Surg. Am.* 75:845-862, 1993.
153. **Ullmark, G.:** Bigger size and defatting of bone chips will increase cup stability. *Arch. Orthop Trauma Surg.* 120:445-447, 2000.
154. **Ullmark, G. and Linder, L.:** Histology of the femur after cancellous impaction grafting using a Charnley prosthesis. *Arch. Orthop Trauma Surg.* 117:170-172, 1998.
155. **Ullmark, G. and Nilsson, O.:** Impacted corticocancellous allografts: recoil and strength. *J Arthroplasty.* 14:1019-1023, 1999.
156. **Ullmark, G. and Obrant, K. J. :** Histology of impacted bone-graft incorporation
2. *J. Arthroplasty.* 17:150-157, 2002.
157. **Urist, M. R.:** Bone: formation by autoinduction. *Science.* 150:893-899, 1965.
158. **Urist, M. R., Dowell, T. A., Hay, P. H., and Strates, B. S.:** Inductive substrates for bone formation. *Clin Orthop.* 59:59-96, 1968.
159. **Urist, M. R., Silverman, B. F., Buring, K., Dubuc, F. L., and Rosenberg, J. M.:** The bone induction principle. *Clin Orthop.* 53:243-283, 1967.
160. **van Biezen, F. C., ten Have, B. L., and Verhaar, J. A.:** Impaction bone-grafting of severely defective femora in revision total hip surgery: 21 hips followed for 41-85 months. *Acta Orthop Scand.* 71:135-142, 2000.
161. **van der, D. S., Buma, P., Slooff, T. J., Gardeniers, J. W., and Schreurs, B. W.:** Incorporation of morselized bone grafts: a study of 24 acetabular biopsy specimens
7. *Clin. Orthop.* 131-141, 2002.
162. **van Doorn, W. J., ten, H. B., van Biezen, F. C., Hop, W. C., Ginai, A. Z., and Verhaar, J. A.:** Migration of the femoral stem after impaction bone grafting. First results of an ongoing, randomised study of the exeter and elite plus femoral stems using radiostereometric analysis. *J. Bone Joint Surg. Br.* 84:825-831, 2002.
163. **van, D., Buma, P., Verdonschot, N., and Schreurs, B. W.:** Effect of load on the early incorporation of impacted morsellized allografts. *Biomaterials.* 23:297-303, 2002.
164. **Verdonschot, N., van Hal, C. T., Schreurs, B. W., Buma, P., Huiskes, R., and Slooff, T. J.:** Time-dependent mechanical properties of HA/TCP particles in relation to morsellized bone grafts for use in impaction grafting. *J Biomed. Mater. Res.* 58:599-604, 2001.
165. **Wang, H. J., Wan, H. L., Yang, T. S., Wang, D. S., Chen, T. M., and Chang, D. M.:** Acceleration of skin graft healing by growth factors. *Burns.* 22:10-14, 1996.

Stimulation and substitution of bone allograft around non-cemented implants

166. **Wang, J. S.:** Basic fibroblast growth factor for stimulation of bone formation in osteoinductive or conductive implants. *Acta Orthop. Scand. Suppl.* 269:1-33, 1996.
167. **Wang, J. S., Tagil, M., and Aspenberg, P.:** Load-bearing increases new bone formation in impacted and morselized allografts. *Clin Orthop.* 274-281, 2000.
168. **Weibrich, G. and Kleis, W. K. :** Curasan PRP kit vs. PCCS PRP system
1. *Clin. Oral Implants. Res.* 13:437-443, 2002.
169. **Weibrich, G., Kleis, W. K., and Hafner, G.:** Growth factor levels in the platelet-rich plasma produced by 2 different methods: curasan-type PRP kit versus PCCS PRP system
4. *Int. J. Oral Maxillofac. Implants.* 17:184-190, 2002.
170. **Whitman, D. H., Berry, R. L., and Green, D. M.:** Platelet gel: an autologous alternative to fibrin glue with applications in oral and maxillofacial surgery. *J Oral Maxillofac Surg.* 55:1294-1299, 1997.
171. **Wozney, J. M.:** Bone morphogenetic proteins. *Prog. Growth Factor. Res.* 1:267-280, 1989.
172. **Xu, J. W., Konttinen, Y. T., Waris, T. F., Lassus, J., Matucci, C. M., Sorsa, T., and Santavirta, T. S.:** Production of platelet-derived growth factor in aseptic loosening of total hip replacement. *Rheumatol. Int.* 17:215-221, 1998.
173. **Yano, H., Ohashi, H., Kadoya, Y., Kobayashi, A., Yamano, Y., and Tanabe, Y.:** Histologic and mechanical evaluation of impacted morselized cancellous allografts in rabbits: comparison with hydroxyapatite granules. *J Arthroplasty.* 15:635-643, 2000.
174. **Zimmermann, R., Jakubietz, R., Jakubietz, M., Strasser, E., Schlegel, A., Wiltfang, J., and Eckstein, R.:** Different preparation methods to obtain platelet components as a source of growth factors for local application. *Transfusion.* 41:1217-1224, 2001.
175. Zohar, R, Oprea, W, Kark, L, Chan, S, and Davies, JE. Mechanism of action of platelet rich plasma on bone healing. *Trans.49th Annual Meeting Orthop Res* , 0493. 2003.
Ref Type: Abstract